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(54) **PROCESS FOR GENERATING SEQUENCE-SPECIFIC PROTEASES BY DIRECTED EVOLUTION**

VERFAHREN ZUR HERSTELLUNG SEQUENZSPEZIFISCHER PROTEASEN DURCH GEZIELTE EVOLUTION

PROCEDE DE PRODUCTION DE PROTEASES A SEQUENCES SPECIFIQUES PAR EVOLUTION DIRIGEE

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(56) References cited:
WO-A-99/11801 US-A1- 2002 022 243

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- **KOLTERMANN ANDRE ET AL: "Principles and methods of evolutionary biotechnology."**
BIOPHYSICAL CHEMISTRY, vol. 66, no. 2-3, 1997,
pages 159-177, XP001085255 ISSN: 0301-4622
- **SMITH T A ET AL: "DIRECT SELECTION FOR SEQUENCES ENCODING PROTEASES OF KNOWN SPECIFICITY" PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF USA, NATIONAL ACADEMY OF SCIENCE.**
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Description

[0001] A process for generating sequence-specific proteases by screening-based directed evolution is disclosed. The use of the process provides proteases recognizing and cleaving user-definable amino-acid sequences with high sequence-specificity. Proteases obtainable by the process can be used in a variety of medical, diagnostic and industrial applications.

Background of the Invention

[0002] Proteolytic enzymes or proteases are a class of enzymes which has an outstanding position among the different enzymes, since the reaction catalyzed by proteases is the cleavage of peptide bonds in other proteins. Proteases are not only very common enzymes in nature, but belong to the most important enzymes for medical and industrial use. Of the total worldwide sales of enzymes, which is estimated to be more than USD 1 billion per year, proteases account for approximately 60 %. Based on the functional group present at the active site, proteases are classified into four groups, i.e., serine proteases (EC 3.4.21), cysteine proteases (EC 3.4.22), aspartic proteases (EC 3.4.23), and metalloproteases (EC 3.4.24). Classification into one of the four groups is typically done by experimental determination of sensitivity towards different types of protease inhibitors. Furthermore, proteases of the four groups differ in their biochemical properties. For example, serine proteases are sensitive to inhibitors 3,4-DCI, DFP, PMSF and TLCK, and have a pH optimum between pH 7 and 11. Aspartic proteases are inhibited by pepstatin, DAN and EPNP, and predominantly have a pH optimum between pH 3 and 4. Cysteine proteases are sensitive to sulfhydryl inhibitors such as PCMB, and besides a few exceptions, have neutral pH optima. Metalloproteases are characterized by the requirement of a divalent metal ion for their activity. Therefore, metalloproteases are inhibited by chelating agents such as EDTA, and have neutral or alkaline pH optima. Among these four groups, further classification is usually done on the basis of structural similarities.

[0003] Besides such a combined biochemical and structural classification, proteases can be grouped according to their substrate spectrum. The two most general groups to be distinguished are exoproteases and endoproteases. Exoproteases only cleave peptide bonds at the very end of an, peptide, whereas endoproteases catalyze the cleavage of bonds anywhere in a peptide strand. The specificity of proteases, i.e. their ability to recognize and hydrolyze specifically certain peptide substrates while others remain uncleaved, can be expressed qualitatively and quantitatively. Qualitative specificity refers to the kind of amino acid residues that are accepted by a protease at certain positions of the peptide substrate. For example, trypsin and the tissue-type plasminogen activator are related with respect to their qualitative specificity, since both of them require at the position P1 an arginine or a similar residue (nomenclature of peptide substrate positions according to the nomenclature of Schlechter & Berger (Biochem. Biophys. Res. Commun. 27 (1967) 157-162). On the other hand, quantitative specificity refers to the relative number of peptide substrates that are accepted as substrates. The quantitative specificity can be expressed by the term

$$s = -\log(Q),$$

where Q is the ratio of all accepted peptide substrates versus all possible peptide substrates. Quantitative specificities of several proteases are shown exemplarily in Table I. The calculation of quantitative specificities is based on the twenty naturally occurring amino acids, and on the assumption that all combinations of these twenty amino acids are feasible. Consequently, proteases that accept only a small portion of all possible peptides have a high specificity, whereas the specificity of proteases that, as an extreme, cleave any peptide substrate would theoretically be zero.

Table I: Quantitative specificities of different proteases

protease	Substrate requirements									Quantitative specificity	
	P6	P5	P4	P3	P2	P1	P1'	P2'	P3'	Q	s = -log Q
	×	×	×	×	×	×	×	×	×	1.00E+00	0
Chymotrypsin	×	×	×	×	×	F/Y/W	×	×	×	1.50E-01	0.82
Papain	×	×	×	×	F/V/L	×	×	×	×	1.50E-01	0.82
Trypsin	×	×	×	×	×	K/R	×	×	×	1.00E-01	1.00

(continued)

protease	Substrate requirements									Quantitative specificity	
	P6	P5	P4	P3	P2	P1	P1'	P2'	P3'	Q	s = -log Q
Pepsin	×	×	×	×	×	F/Y/L	W/F/Y	×	×	2.25E-02	1.65
TEV	E	×	×	Y	×	Q	S/G	×	×	1.25E-05	4.90
Plasmin	×	×	K/V/I/F	×	F/Y/W	R/K	N	A	×	7.50E-06	5.12
thrombin	×	×	L/I/V/F	×	P	R	N	A	×	1.25E-06	5.90
t-PA	×	×	C	P	G	R	V	V	G	7.81E-10	9.11
(Amino acid residues are abbreviated as shown in Table II. X refers to any amino acid residue.)											

[0004] The quantitative specificity of proteases varies over a wide range. There are very unspecific proteases known, such as papain which cleaves all polypeptides that contain a phenylalanine, a valine or an leucine residue ($s = 0.82$), or trypsin which cleaves all polypeptides that contain an arginine or a lysine residue ($s = 1.0$). On the other hand, there are highly specific proteases known, such as the tissue-type plasminogen activator (t-PA) which cleaves plasminogen only at a single specific sequence ($s = 9.11$). Proteases with high substrate specificity play an important role in the regulation of protein functions in living organisms. The specific cleavage of polypeptide substrates, for example, activates precursor proteins or deactivates active proteins or enzymes, thereby regulating their functions. Several proteases with high substrate specificities are used in medical applications. Pharmaceutical examples for activation or deactivation by cleavage of specific polypeptide substrates are the application of t-PA in acute cardiac infarction which activates plasminogen to resolve fibrin clots, or the application of Ancrod in stroke which deactivates fibrinogen, thereby decreasing blood viscosity and enhancing its transport capacity. While t-PA is a human protease with an activity necessary in human blood regulation, Ancrod is a non-human protease. It was isolated from the viper *Agkistrodon rhodostoma*, and comprises the main ingredient of the snake's poison. Therefore, there exist a few non-human proteases with therapeutic applicability. Their identification, however, is usually highly incidental.

[0005] The treatment of diseases by administering drugs is typically based on a molecular mechanism initiated by the drug that activates or inactivates a specific protein function in the patient's body, be it an endogenous protein or a protein of an infecting microbe or virus. While the action of chemical drugs on these targets is still difficult to understand or to predict, protein drugs are able to specifically recognize these target proteins among millions of other proteins. Prominent examples of proteins that have the intrinsic possibility to recognize other proteins are antibodies, receptors, and proteases. Although there are a huge number of potential target proteins, only very few proteases are available today to address these target proteins. Due to their proteolytic activity, proteases are particularly suited for the inactivation or activation of protein targets. When considering human proteins only, the number of potential target proteins is yet enormous. It is estimated that the human genome comprises between 30,000 and 100,000 genes, each of which encodes a different protein. Many of these proteins are involved in human diseases and are therefore potential pharmaceutical targets. Proteases recognizing and cleaving these target proteins with a high specificity are consequently of high value as potential drugs. The medical application of such proteases, however, is restricted by their occurrence. For example, there are theoretically 25 billion different possibilities for a specificity of $s = 10.4$ (corresponding to the specific recognition of a unique sequence of eight amino acid residues). It is highly unlikely to find such a protease with one particular qualitative specificity by screening natural isolates.

[0006] Selection systems for proteases of known specificity are known in the art, for instance, from Smith et al., Proc. Natl. Acad. Sci. USA, Vol. 88 (1991). As exemplified, the system comprises the yeast transcription factor GAL4 as the selectable marker, a defined and cleavable target sequence inserted into GAL4 in conjunction with the TEV protease. The cleavage separates the DNA binding domain from the transcription activation domain and therewith renders the transcription factor inactive. The phenotypical inability of the resulting cells to metabolize galactose can be detected by a colorimetric assay or by the selection on the suicide substrate 2-deoxygalactose.

[0007] Further, selection may be performed by the use of peptide substrates with modifications as, for example, fluorogenic moieties based on groups as ACC, previously described by Harris et al. (US 2002/022243).

[0008] Laboratory techniques to generate proteolytic enzymes with altered sequence specificities are in principle known. They can be classified by their expression and selection systems. Genetic selection means to produce a protease within an organism which protease is able to cleave a precursor protein which in turn results in an alteration of the growth behavior of the producing organism. From a population of organisms with different proteases those having an altered growth behavior can be selected. This principle was reported by Davis et al. (US 5258289, WO 96/21009). The production

of a phage system is dependent on the cleavage of a phage protein which only can be activated in the presence of a proteolytic enzyme or antibody which is able to cleave the phage protein. Selected proteolytic enzymes or antibodies would have the ability to cleave an amino acid sequence for activation of phage production. Furthermore, there is no control of the specificity of the proteases that are selected. The system does not select for proteases with low activities for other peptides than the used peptide substrate. Additionally, this system does not allow a precise characterization of the kinetic constants of the selected proteases (k_{cat} , K_M). Several other systems with intracellular protease expression are reported but they all suffer from the disadvantages mentioned above. Some of them use a genetic reporter system which allows a selection by screening instead of a genetic selection, but also cannot overcome the intrinsic insufficiency of the intracellular characterization of proteases.

[0009] A system to generate proteolytic enzymes with altered sequence specificities with membrane-bound proteases is reported. Iverson et al. (WO 98/49286) describe an expression system for a membrane-bound protease which is displayed on the surface of cells. An essential element of the experimental design is that the catalytic reaction has to be performed at the cell surface, i.e., the substrates and products must remain associated with the bacterium expressing the enzyme at the surface. This restriction limits the generation of proteolytic enzymes with altered sequence specificities and does not allow a precise characterization of the kinetic constants of the selected proteases (k_{cat} , K_M). Furthermore, the method does not allow the control of the position at which the peptide is cleaved. Additionally, positively identified proteases will have the ability to cleave a certain amino acid (aa) sequence but they also may cleave many other aa sequences. Therefore, there is no control of the specificity of the proteases that are selected.

[0010] A system to generate proteolytic enzymes with altered sequence specificities with self-secreting proteases is also known. Duff et al. (WO 98/11237) describe an expression system for a self-secreting protease. An essential element of the experimental design is that the catalytic reaction acts on the protease itself by an autoproteolytic processing of the membrane-bound precursor molecule to release the matured protease from the cellular membrane into the extracellular environment. Therefore, a fusion-protein must be constructed where the target peptide sequence replaces the natural cleavage site for autoproteolysis. Limitations of such a system are that positively identified proteases will have the ability to cleave a certain aa sequence but they also may cleave many other peptide sequences. Therefore, high substrate specificity cannot be achieved with such an approach. Additionally, such a system is not able to control that selected proteases cleave at a specific position in a defined aa sequence and it does not allow a precise characterization of the kinetic constants of the selected proteases (k_{cat} , K_M).

[0011] Broad et al. (WO 99/11801) disclose a heterologous cell system suitable for the alteration of the specificity of proteases. The system comprises a transcription factor precursor wherein the transcription factor is linked to a membrane anchoring domain via a protease cleavage site. The cleavage at the protease cleavage site by a protease releases the transcription factor, which in turn initiates the expression of a target gene being under the control of the respective promoter. The experimental design of alteration of the specificity consists in the insertion of protease cleavage sites with modified sequences and the subjection of the protease to mutagenesis. New proteases obtained may be able to recognize the modified sequence, the effect of which is monitored by the expression of the target gene. Such a system does also not allow a precise control of biochemical properties of the selected proteases.

[0012] Most of these approaches apply methods of directed evolution for the generation of proteolytic enzymes with altered sequence specificities. Several different mutation and recombination methods to generate genetic libraries are reported and described elsewhere. All the different methods suffer from their lack of precise selection of positive protease variants from large libraries. First, these methods are not able to distinguish between single and multi turn-overs of peptide substrates which is necessary in order to prevent the selection of low k_{cat} variants. Secondly, it is not possible to trigger enzyme and substrate concentration to select protease variants for lower K_M . Third, none of these systems allows the selection of a protease with an increased activity on the desired peptide substrate whereby the activity on the original peptide substrate decreases.

[0013] Methods which fulfill the above mentioned three selection criteria (k_{cat} , K_M and substrate specificity) for generating proteolytic enzymes with high sequence-specificity applying screening-based directed evolution have heretofore not been available.

Summary of the Invention

[0014] Thus, the technical problem underlying the present invention is to provide a method for generating new proteases with user-defined substrate specificities by applying directed evolution. In particular, the invention is directed to a method for the evolution of novel proteases towards selective recognition and cleavage of specific amino-acid sequences only. This technical problem has been solved by the embodiments of the invention specified below and in the appended claims. The present invention is thus directed to

- (1) a method for identifying sequence-specific proteases with target substrate specificities which comprises the following steps

(a) providing a population of proteases comprised of variants of one first protease or of variants or chimeras of two or more first proteases, said first proteases having a substrate specificity for a particular amino acid sequence of a first peptide substrate;

(b) contacting said population of proteases with one or more second substrates, comprising at least one specific amino acid sequence resembling the amino acid sequence of the target peptide substrate but being not present within the first peptide substrate; and

(c) selecting one or more protease variants from the population of proteases provided in step (a) having specificity for said specific amino acid sequence of the second substrates provided in step (b) under conditions that allow identification of proteases that recognize and hydrolyse preferably said specific one amino acid sequence within the second substrates, wherein the screening for protease activity is achieved by adding in excess peptides other than the second peptide; thereby using the added peptides as competitors;

(2) in a preferred embodiment of (1) above only one second substrate is used in the one or more cycles (a) to (c), i.e., the second substrate is identical with the target substrate;

(3) in a further preferred embodiment of (1) above different second substrates are used, and the second substrates have an intermediate character with regard to the first substrate and the target substrate, and the last second substrate that is used is identical with the target substrate; and

(4) in a particular preferred embodiment of (1) to (3) above the target protease has a specificity similar to tissue-type plasminogen activator and cleaves the target substrate CPGR[↓]VVGG.

[0015] The identification and selection of proteases that have evolved towards the target specificity is done by screening for catalytic activities on different peptide substrates, either by screening for increased affinity, or by using two substrates in comparison, or by using unspecific peptides as competitors, or by using intermediate peptide substrates. The following detailed description will disclose the preferred features, advantages and the utility of the present invention.

Brief Description of the Figures

[0016] The following figures are provided in order to explain further the present invention in supplement to the detailed description:

- Figure 1 depicts schematically the two alternatives A and B of the method of the invention.
- Figure 2 distinguishes the two alternatives A and B of the method of the invention by showing schematically the qualitative and quantitative changes in specificity during evolution towards the target specificity.
- Figure 3 illustrates schematically how proteases with changed catalytic activities are evolved using the two alternatives A and B of the method of the invention.
- Figure 4 depicts schematically in two different forms the intermediate approach as one particular aspect of the invention that uses intermediate substrates.
- Figure 5 illustrates schematically how, according to the invention, proteases with changed catalytic activities are evolved using the intermediate approach.
- Figure 6 shows exemplarily an expression vector for *S. cerevisiae* that can be used for the method of the invention.
- Figure 7 shows exemplarily the hydrolysis of a peptide substrate by the tobacco etch virus protease.
- Figure 8 shows exemplarily a distribution of catalytic activities obtained by screening using confocal fluorescence spectroscopy.
- Figure 9 shows exemplarily the decrease in K_M during evolution towards higher affinity.
- Figure 10 shows exemplarily the change in specificity during evolution of proteases towards the specificity of t-PA.
- Figure 11 depicts schematically a preferred variant of the intermediate approach.
- Figure 12 shows exemplarily the time-dependent substrate conversion of a starting protease in comparison to one of the evolved variants.

Detailed Description of the Invention

[0017] In the framework of this invention the following terms and definitions are used.

[0018] The term "protease" means any protein molecule acting in the hydrolysis of peptide bonds. It includes naturally-occurring proteolytic enzymes, as well as variants thereof obtained by site-directed or random mutagenesis or any other protein engineering method, any fragment of a proteolytic enzyme, or any molecular complex or fusion protein comprising one of the aforementioned proteins. A "chimera of proteases" means a fusion protein out of two or more fragments derived from different parent proteases.

[0019] The term "substrate" or "peptide substrate" means any peptide, oligopeptide, or protein molecule of any amino

acid composition, sequence or length, that contains a peptide bond that can be hydrolyzed catalytically by a protease. The peptide bond that is hydrolyzed is referred to as the "cleavage site". Numbering of positions in the substrate is done according to the system introduced by Schlechter & Berger (Biochem. Biophys. Res. Commun. 27 (1967) 157-162). Amino acid residues adjacent N-terminal to the cleavage site are numbered P1, P2, P3, etc., whereas residues adjacent C-terminal to the cleavage site are numbered P1', P2', P3', etc.

[0020] The term "specificity" means the ability of a protease to recognize and hydrolyze selectively certain peptide substrates while others remain uncleaved. Specificity can be expressed qualitatively and quantitatively. "Qualitative specificity" refers to the kind of amino acid residues that are accepted by a protease at certain positions of the peptide substrate. "Quantitative specificity" refers to the number of peptide substrates that are accepted as substrates. Quantitative specificity can be expressed by the term s , which is the negative logarithm of the number of all accepted peptide substrates divided by the number of all possible peptide substrates. Proteases that accept only a small portion of all possible peptide substrates have a "high specificity" ($s \gg 1$). Proteases that accept almost any peptide substrate have a "low specificity". Proteases with very low specificity ($s \leq 1$) are also referred to as "unspecific proteases".

[0021] The term "first protease" describes any protease used in step (a) of this invention as the starting point in order to generate populations of protease variants that are related to this first protease. The term "first substrate" or "first peptide substrate" describes a substrate that is recognized and hydrolyzed by the first protease. The term "first specificity" describes the qualitative and quantitative specificity of the first protease.

[0022] The term "evolved protease" describes any protease that is generated by use of the method of the invention. The term "target substrate" or "target peptide substrate" describes a substrate that is recognized and hydrolyzed by the evolved protease. The term "target specificity" describes the qualitative and quantitative specificity of the evolved protease that is to be generated by use of the method of the invention. Thus, the target specificity defines the specificity of the evolved protease for the target peptide substrate while other substrates are not or very weakly recognized and hydrolyzed.

[0023] The term "intermediate" or "intermediate substrate" describes any substrate that has an intermediate character between two other substrate. The intermediate character can base on the amino acid composition, the amino acid sequence, the properties of the amino acid residues contained in the substrates, or a combination of these characteristics.

[0024] Catalytic properties of proteases are expressed using the kinetic parameters " K_M " or "Michaelis Menten constant", " k_{cat} " or "catalytic rate constant", and " k_{cat}/K_M " or "catalytic efficiency", according to the definitions of Michaelis and Menten (Fersht, A., Enzyme Structure and Mechanism, W. H. Freeman and Company, New York, 1995). The term "catalytic activity" describes the rate of conversion of the substrate under defined conditions.

[0025] Amino acids are abbreviated according to the following Table II either in one- or in three-letter code.

Table II: Amino acid abbreviations

Abbreviations		Amino acid
A	Ala	Alanin
C	Cys	Cysteine
D	Asp	Aspartic cid
E	Glu	Glutamic acid
F	Phe	Phenylalanine
G	Gly	Glycine
H	His	Histidine
I	Ile	Isoleucine
K	Lys	Lysine
L	Leu	Leucine
M	Met	Methionine
N	Asn	Asparagine
P	Pro	Proline
Q	Gin	Glutamine
R	Arg	Arginine
S	Ser	Serine

(continued)

Abbreviations		Amino acid
T	Thr	Threonine
V	Val	Valine
W	Trp	Tryptophane
Y	Tyr	Tyrosine

[0026] As set forth above, the present invention is directed to a method for generating sequence-specific proteases with a target substrate specificity by applying principles of molecular evolution. According to the invention, this is achieved by providing a population of proteases being related to each other, as well as a peptide substrate that resembles the target substrate, and selecting one or more protease variants from the population of proteases with respect to their specificity for the provided substrate. The selection is done under conditions that allow identification of proteases that recognize and hydrolyze the target sequence preferably.

[0027] In particular, embodiment (1) of the invention relates to a method for generating sequence-specific proteases with target substrate specificities, wherein the following steps are carried out:

- (a) providing a population of proteases, wherein each variant is related to one or more first proteases, these first proteases having a first substrate specificity;
- (b) providing one or more peptide substrates comprising at least one amino-acid sequence that resembles the target peptide substrate;
- (c) selecting one or more protease variants from the population of proteases provided in step (a) with respect to their specificity for the substrate provided in step (b) under conditions that allow identification of proteases that recognize and cleave the target sequence preferably;

and wherein steps (a) to (c) are carried out cyclically until one or more protease variants with the target substrate specificity are identified.

[0028] When repeating steps (a) to (c), the one or more proteases selected in step (c) of one cycle are used as the one or more first proteases in step (a) of the next cycle.

[0029] In one alternative of the invention, the one or more first proteases serving as starting points in step (a) of the method have a high sequence specificity which is maintained high during the directed evolution towards the target specificity.

[0030] In another alternative of the method, the one or more first proteases serving as starting points in step (a) of the method have a low sequence specificity, which is increased during the directed evolution towards the target specificity.

[0031] The steps (a) to (c) of the above method are carried out for at least one cycle. Preferably, however, these steps are carried out for several cycles, with each one or more protease variants selected in one cycle being the origin of the population of protease variants in the next cycle. Preferably, more than one and less than hundred, more preferably more than two and less than fifty, particularly preferably more than three and less than twenty, especially preferably more than four and less than ten, and most preferably five cycles of steps (a) to (c) are carried out until one or more protease variants with the target substrate specificity are identified.

[0032] The invention applies evolutionary means as described in very detail in WO9218645 with that document being incorporated in its entirety for all purposes.

[0033] For an overview on the application of evolutionary principles to molecular biotechnology, which is usually referred to as "directed evolution" or "evolutionary biotechnology", see the review by Koltermann & Kettling (Biophys. Chem. 66 (1997) 159-177).

[0034] Part of the invention is the provision of populations of protease variants wherein each variant is related to one or more first proteases. In principle, there can be a large number of these first proteases, all together being the origin for the first cycle of the method. It is preferred, however, that these first proteases comprise fifty or less different proteases, more preferably ten or less different proteases, especially preferably two or less different proteases. Most preferably, only one first protease is employed.

[0035] According to the invention, any protease can be used as first protease. Preferably, an endoprotease is used as first protease. It is preferred that the protease belongs to the group of proteases consisting of Serine proteases (EC 3.4.21), Cysteine proteases (EC 3.4.22), Aspartic proteases (EC 3.4.23), and Metalloproteases (EC 3.4.24). First proteases are characterized by their ability to recognize and hydrolyze peptide substrates with a certain qualitative and quantitative specificity. First proteases can have a specificity in the same range as the specificity of the protease that is to be generated. Examples for proteases with relatively high specificities are TEV protease, HIV-1 protease, BAR1

protease, Factor Xa, Thrombin, tissue-type plasminogen activator, Kex2 protease, TVMV-protease, RSV protease, MuLV protease, MPMV protease, MMTV protease, BLV protease, EIAV protease, SIVmac protease. Alternatively, the first proteases have a lower specificity than the specificity of the protease that is to be generated. As an extreme example of the latter, proteases with very low sequence specificity are employed, for example proteases such as Papain, Trypsin, Chymotrypsin, Subtilisin, SET (trypsin-like serine protease from *Streptomyces erythraeus*), Elastase, Cathepsin G or Chymase.

[0036] A particularly suitable protease is sp |P12630 | BAR1 protease (BAR1_YEAST Barrierpepsin precursor (EC 3.4.23.35) (extracellular "barrier" protein) (BAR proteinase) of *S. cerevisiae* (see SEQ ID NO:8).

[0037] The provision of populations of proteases is essentially done as described in WO9218645. According to the invention, genes encoding protease variants are ligated into a suitable expression vector by standard molecular cloning techniques (Sambrook, J.F; Fritsch, E.F.; Maniatis, T.; Cold Spring Harbor Laboratory Press, Second Edition, 1989, New York). The vector is introduced in a suitable expression host cell, which expresses the corresponding protease variant. Particularly suitable expression hosts are bacterial expression hosts such as *Escherichia coli* or *Bacillus subtilis*, or yeast expression hosts such as *Saccharomyces cerevisiae* or *Pichia pastoris*, or mammalian expression hosts such as Chinese Hamster Ovary (CHO) or Baby Hamster Kidney (BHK) cell lines, or viral expression systems such as the Baculovirus system. Alternatively, systems for *in vitro* protein expression can be used.

[0038] In a preferred embodiment of the invention, the genes are ligated into the expression vector behind a suitable signal sequence that leads to secretion of the protease variants into the extracellular space, thereby allowing direct detection of protease activity in the cell supernatant. Particularly suitable signal sequences for *Escherichia coli* are HlyA, for *Bacillus subtilis* AprE, NprB, Mpr, AmyA, AmyE, Blac, SacB, and for *S. cerevisiae* Bar1, Suc2, Mata, Inu1A, Ggplp.

[0039] In another preferred embodiment of the invention, the protease variants are expressed intracellularly and the peptide substrates are expressed also intracellularly. Preferably, this is done essentially as described in WO 0212543, using a fusion peptide substrate comprising two auto-fluorescent proteins linked by the substrate amino-acid sequence.

[0040] In another preferred embodiment of the invention, the protease variants are expressed intracellularly, or secreted into the periplasmic space using signal sequences such as DsbA, PhoA, PelB, OmpA, OmpT or gIII for *Escherichia coli*, followed by permeabilisation or lysis step to release the protease variants into the supernatant. The destruction of the membrane barrier can be forced by the use of mechanical means such as ultrasonic, French press, or the use of membrane-digesting enzymes such as lysozyme.

[0041] As a further alternative, the genes encoding the protease variants are expressed cell-free by the use of a suitable cell-free expression system. In a particularly preferred embodiment, the S30 extract from *Escherichia coli* cells is used for this purpose as described by Lesly et al. (Methods in Molecular Biology 37 (1995) 265-278).

[0042] The relatedness to the one or more first proteases can be achieved by several procedures. For example, the genes encoding the one or more first proteases are modified by methods for random nucleic acid mutagenesis. In a preferred embodiment of the invention, random mutagenesis is achieved by the use of a polymerase as described in WO 9218645. According to this embodiment, the one or more genes encoding the one or more first proteases are amplified by the use of a polymerase with a high error rate, or under conditions that increase the rate of misincorporations, thereby leading to a population of genes wherein each gene encodes a protease that is related to the one or more first proteases. For example the method according to Cadwell, R.C and Joyce, G.F. can be employed (PCR Methods Appl. 2 (1992) 28-33). Other methods for random mutagenesis that can be employed make use of mutator strains, UV-radiation or chemical mutagens. Most preferably, errors are introduced into the gene at or near but below the error threshold as described in WO 9218645.

[0043] In another preferred embodiment of the invention, certain parts of the gene encoding the protease variants are randomized completely with respect to the amino-acid sequence, and are re-introduced into the gene as an oligonucleotide cassette. This technique is usually referred to as cassette mutagenesis (Oliphant, A.R. et al., Gene 44 (1986) 177-183; Horwitz, M.S., et al. Genome 31 (1989) 112-117). In a particularly preferred embodiment of the invention, the part of the gene that encodes amino acid residues that are essential for recognition of the substrate is randomized via cassette mutagenesis. These residues can be identified from structural studies. In particular, residues comprising parts of the substrate binding pocket are targeted by cassette mutagenesis. Alternatively, substituting each amino acid residue with an alanine, and analyzing whether there is an effect on the catalytic activity can identify such residues. As a further alternative, these residues can be identified by first introducing random mutations into the gene, screening for an effect on specificity, affinity, or catalytic activity, and determining afterwards the position of mutations in variants that represent altered specificity, affinity or altered catalytic activity. As an extreme of this approach, the completely randomized sequence can have the length of one nucleotide only. This approach is typically referred to as site saturation mutagenesis.

[0044] In another preferred embodiment of the invention, nucleic acid sequences are randomly introduced into or deleted from the one or more first protease genes in order to provide a population of proteases. This approach is referred to as insertion and/or deletion mutagenesis. For insertion mutagenesis, random sequences of defined or random length are introduced randomly into a gene. As an example, the method described by Hallet et al. (Nucleic Acids Res. 1997, vol. 25, p. 1866ff) can be used to introduce a random 15 nt sequence randomly into a gene. Alternatively, defined

sequences, for example a sequence encoding a specific protein secondary structure motif, can be inserted randomly into a gene. Alternatively, random sequences of defined or random length can be inserted at specific sites into a gene. This can be done using restriction sites or by oligonucleotide overlap extension methods such as the method described by Horton (Gene 1989, vol. 77, p. 61ff). For deletion mutagenesis, sequences of defined or random length are deleted randomly from a gene. In a particular embodiment of the invention, deletion and insertion mutagenesis are combined so that insertions at one site can potentially be combined, and thereby possibly compensated, by deletion at another site.

[0045] In a further preferred embodiment of the invention, methods for homologous in-vitro recombination are used for the provision of protease populations. Examples of methods that can be applied are the Recombination Chain Reaction (RCR) according to WO 0134835, the DNA-Shuffling method according to WO 9522625, the Staggered Extension method according to WO 9842728, or the Random Priming recombination according to WO9842728. Furthermore, also methods for non-homologous recombination such as the Itchy method can be applied (Ostermeier, M. et al., Nature Biotechnology 17 (1999) 1205-1209). All of the references mentioned above are hereby incorporated by reference in its entirety for all purposes.

[0046] In further embodiments of the invention, the above-mentioned methods are combined with each other. In a particularly preferred embodiment, the Recombination Chain Reaction is combined with random mutagenesis such as error-prone PCR according to Cadwell, R.C and Joyce, G.F. (PCR Methods Appl. 2 (1992) 28-33) in order to de-couple mutations selected in the round before and to introduce simultaneously a defined number of new random mutations into the population.

[0047] The coupling of protease genotype and phenotype is achieved by use of sample carriers that enable compartmentation of samples, and the distribution of genotypes into sample carriers is done at a multiplicity per compartment that allows sufficient differentiation of phenotypes.

[0048] The one or more first proteases that serve as the starting point of the method either have a specificity which is in the range of the target specificity that is to be generated by the method, or have a lower specificity than the target specificity. Accordingly, the method of the invention is either performed under conditions that maintain the specificity quantitatively and alters it qualitatively (Alternative A), or the method of the invention is performed under conditions that maintains the specificity qualitatively and increases it quantitatively (Alternative B). Moreover, both approaches can be combined. These three principle alternatives are shown schematically in Figure 2.

[0049] In a preferred embodiment of the invention corresponding to alternative A, the one or more first proteases have a first specificity that is quantitatively in the range of the target specificity, but qualitatively distinct from the target specificity. Proteases having the target substrate specificity are achieved using the method of the invention by selecting protease variants under conditions that allow identification of proteases that recognize and cleave the target sequence preferably.

[0050] In another preferred embodiment of the invention corresponding to alternative B, the specificity of the one or more first proteases is quantitatively lower when compared to the target specificity. This means that they accept and hydrolyze a larger number of peptide substrates. This low first specificity is subsequently

[0051] increased by the method of the invention until it is in the range of the target specificity. As a preferred variant of this embodiment, the first specificity is qualitatively related to the target specificity. Thus, the large number of peptide substrates that is accepted and hydrolyzed includes the target substrate already. Accordingly, amino acid residues that are essential in the first substrate remain essential residues in the target substrate. Then, proteases having the target substrate specificity are achieved using the method of the invention by selecting protease variants under conditions that allow identification of proteases that recognize and cleave the target sequence preferably.

[0052] Another part of the invention is the provision of peptide substrates that resemble the target substrate, and the use of these substrates for screening of protease variants with respect to their catalytic activity.

[0053] In a preferred embodiment of the invention, suitable peptide substrates are synthesized via the solid phase peptide synthesis approach of Merrifield et al (Nature. 207 (1965) 522-523). These peptide substrates are then incubated for a certain time in a sample buffer containing the protease variant to be tested. The hydrolysis of the peptide is then analyzed by a suitable method. For example, the amount of fragmented peptides can be analyzed by chromatography. In particular, peptide fragments are analyzed advantageously on a reversed phase HPLC system. Alternatively, the peptide substrate is modified in any way to enable the analysis of peptide hydrolysis. In particular, the peptide substrate may carry functional groups that enable the detection of the hydrolysis of the substrate. Such functional groups include, but are not limited to, the following:

- one or more fluorophores or chromophores, whose spectroscopic properties change upon hydrolysis of the peptide, whereby screening is performed through determination of the change in spectroscopic properties; or
- two fluorophores which are distinguishable by their fluorescence properties and which are attached to opposite ends of the second substrate, whereby the screening is performed through confocal fluorescence spectroscopy at fluorophore concentrations below 1 μ M; or
- two fluorophores which form a fluorescence resonance energy transfer (FRET) pair and which are attached to opposite ends of the second substrate, whereby screening is performed through determination of the decrease in

the energy transfer between the two fluorophores; or
 a first and second autofluorescent protein flanking the second substrate, whereby the screening is performed through
 confocal fluorescence spectroscopy at substrate concentrations below 1 μ M; or
 a fluorophore and a quencher molecule which are attached to opposite ends of the second substrate, whereby
 5 screening is performed through determination of the decrease in quenching of the fluorophore; or
 a fluorophore or a chromophore and a binding moiety which are attached to opposite ends of the second substrate,
 whereby screening is performed through determination of binding of the binding moiety to a specific binding partner ;
 or
 a radioactive label and a binding moiety which are attached to opposite ends of the second substrate, whereby
 10 screening is performed through use of a scintillation proximity assay; or
 any combination thereof.

[0054] With respect to the above mentioned functional groups, a chemical group can be attached to the peptide that
 alters its properties when the peptide is hydrolyzed. For example, a para-nitrophenyl group can be used for this purpose.
 15 As another example, one or more fluorophores and/or a quencher molecule are attached to the peptide, and the amount
 of fragmented peptide is analysed by measuring a difference in the fluorescence of the fluorophores. For example, two
 fluorophores that are suited to form a FRET (fluorescence resonance energy transfer) pair are attached to the peptide
 at opposite ends, and the hydrolysis of the peptide is measured by a decrease in the energy transfer between the two
 fluorophores. For example, Rhodamine Green (Molecular Probes Inc., Oregon, USA) and Tetramethylrhodamine (Mo-
 20 lecular Probes Inc., Oregon, USA) can be used as fluorophores that are suited to form such a FRET pair.

[0055] In a particularly preferred embodiment of the invention, two fluorophores that do not form a substantial FRET
 pair are attached to opposite ends of synthetic peptide substrates. As an example, Rhodamine Green (Molecular Probes
 Inc., Oregon, USA) and Cy-5 (Amersham Biosciences Europe GmbH, Freiburg) can be used for this purpose, and
 covalent attachment of the dye can be achieved via a succinimidyl ester linkage to a primary amino group of the peptide.
 25 Hydrolysis of these peptides is preferably analysed by means of confocal fluorescence spectroscopy according to patent
 applications WO 9416313 and WO9613744, which are hereby incorporated by reference in their entirety for all purposes.
 Due to the high sensitivity of confocal fluorescence spectroscopy, substrates are used in concentrations below one
 micromolar, more preferably below hundred nanomolar, and most preferably below ten nanomolar. Therefore, screening
 according to this embodiment is done substantially below the K_M of typical proteases.

[0056] In another particularly preferred embodiment of the invention, fusion proteins comprising a first autofluorescent
 protein, a peptide, and a second autofluorescent protein are used as peptide substrates. According to WO0212543,
 which is hereby incorporated by reference in its entirety for all purposes, autofluorescent include the Green Fluorescent
 Protein GFP and its mutants, as well as dsRED and its mutants. Fusion proteins can be produced by expression of a
 suitable fusion gene in *E. coli*, lysis of cells and purification of the fusion protein by standard methods such as ion
 35 exchange chromatography or affinity chromatography.

[0057] It is an essential part of the invention that proteases with the target substrate specificity are generated by
 selecting protease variants under conditions that allow identification of proteases that recognize and cleave the target
 sequence preferably. This selection can be achieved according to the different aspects of the invention as outlined below.

[0058] In a first aspect of the invention, proteases that recognize and cleave the target sequence preferably are
 40 identified by screening for proteases with a high affinity for the target substrate sequence. High affinity corresponds to
 a low K_M which is selected by screening at target substrate concentrations substantially below the K_M of the first protease.
 This aspect is referred to as the "affinity approach".

[0059] In a preferred embodiment of this aspect of the invention, the peptide substrate provided in step (b) is linked
 to one or more fluorophores that enable the detection of the hydrolysis of the peptide substrate at concentrations below
 45 10 μ M, preferably below 1 μ M, more preferably below 100 nM, and most preferably below 10 nM.

[0060] In a second aspect of the invention, proteases that recognize and cleave the target sequence only are identified
 by providing two or more peptide substrates in step (b) and by screening for activity on these two or more peptide
 substrates in comparison. This aspect is referred to as the "comparison approach".

[0061] In a preferred embodiment of this aspect of the invention, the two or more peptide substrates provided in step
 50 (b) are linked to different marker molecules, thereby enabling the detection of the cleavage of the two or more peptide
 substrates consecutively or in parallel. In a particularly preferred embodiment of the invention, two peptide substrates
 are provided in step (b), one peptide substrate having an amino acid sequence identical to or resembling the first peptide
 substrate thereby enabling to monitor the original activity of the first proteases, and the other peptide substrate having
 an amino-acid sequence identical to or resembling the target substrate sequence thereby enabling to monitor the activity
 55 on the target substrate. In an especially preferred embodiment of the invention, these two peptide substrates are linked
 to fluorescent marker molecules, and the fluorescent properties of the two peptide substrates are sufficiently different
 in order to distinguish both activities when measured consecutively or in parallel. For example, a fusion protein comprising
 a first autofluorescent protein, a peptide, and a second autofluorescent protein according to patent application WO

0212543 can be used for this purpose. Alternatively, fluorophores such as rhodamines are linked chemically to the peptide substrates.

[0062] In a third aspect of the invention, proteases that recognize and cleave the target sequence preferably are identified by providing in step (b) one or more peptide substrates resembling the target peptide together with competing peptide substrates in high excess. Screening with respect to activity on the substrates resembling the target substrate is then done in the presence of the competing substrates. Proteases having a specificity which corresponds qualitatively to the target specificity, but having only a low quantitative specificity are identified as negative samples in such a screen. Whereas proteases having a specificity which corresponds qualitatively and quantitatively to the target specificity are identified positively. This aspect is referred to as the "competitor approach".

[0063] In a preferred embodiment of this aspect of the invention, the one or more peptide substrates resembling the target substrate are linked to marker molecules, thereby enabling the detection of their hydrolysis, whereas the competing peptide substrates do not carry marker molecules. The competing peptide substrates have an amino-acid sequence identical to or resembling the first peptide substrate, or have random amino-acid sequences, thereby acting as competitive inhibitors for the hydrolysis of the marker-carrying peptide substrates.

[0064] In a fourth aspect of the invention, proteases that recognize and cleave the target sequence preferably are identified by using intermediate substrates for evolving the protease towards the target substrate specificity. This aspect is hereinafter also referred to as the "intermediate approach". In a first variant of this aspect of the invention, this is achieved by providing in different cycles different peptide substrates, whereby each peptide substrate has an intermediate character with regard to the cycle before and the target peptide substrate. According to this variant, proteases are evolved gradually toward the target specificity. Figure 4 depicts schematically the basic principle of this variant of the intermediate approach.

[0065] More generally, a first variant of this aspect of the invention is directed to a method for generating sequence-specific proteases with a target substrate specificity, wherein the following steps are carried out:

- (a) providing a population of proteases, wherein each variant is related to one or more first proteases, these first proteases having specificity for a spectrum of peptide substrates or a single peptide substrate;
- (b) providing one or more peptide substrates that has an intermediate character with regard to the first peptide substrate and the target substrate;
- (c) selecting one or more protease variants from the population of proteases provided in step (a) with respect to their specificity for the substrate provided in step (b);
- (d) repeating steps (a) to (c) until one or more protease variants with activity for the intermediate substrate provided in step (b) are identified;
- (e) replacing the first peptide substrate in steps (a) and (b) with the intermediate substrate, and the first proteases in step (a) with the protease variants selected in step (c);

and repeating steps (a) to (e) until one or more protease variants with the target substrate specificity are identified.

[0066] In this first variant of this aspect of the invention, evolution of protease specificity is directed via consecutive selection on a certain number of intermediate peptide substrates, whereby every peptide substrate resembles more and more the target peptide sequence. This approach is based on the finding that proteases which accept related substrates are usually also related to each other. Relatedness of proteases in the context of this invention is a measure for the homology in the amino acid sequences of two or more enzymes. Moreover, this approach is based on the surprising discovery, that distinguishable subsites in a protease active site can be evolved separately, and that their molecular structure can be attributed to different residues of a peptide substrate (Schlechter & Berger, Biochem. Biophys. Res. Commun. 27 (1967) 157-162).

[0067] Intermediate substrates can be realized by substituting amino acid residues at one or more positions from the first peptide sequence with amino acid residues at the same positions from the target peptide sequence. Such intermediates are referred to as "amino acid composition intermediates". Additionally, an intermediate peptide substrate can include one or more amino acid residues at one or more positions which are neither the residues of the first peptide sequence nor the residues of the target peptide sequence at that position, but are amino acid residues with an intermediate character with respect to the residues in the first and the target substrate. Such intermediates are referred to as "amino acid property intermediates". The intermediate character of this kind of intermediates can be based on one or more physical and chemical parameters, which include, but are not limited to, the surface of the residue, its volume, the isoelectric point, the side chain pKa, the polarity, the ability to form hydrogen bonds or the hydrophobicity. In the following table, the twenty naturally occurring amino acid residues are classified according to these parameters.

Table III: Classification of the 20 naturally occurring amino acid residues

Amino acid residue		Type	Surface ^a [Å ²]	Volume ^b [Å ³]	Side chain pK _a ^c (charge) ^d	Relative Hydrophobicity ^e	Hydrogen bond donor or acceptor	
5	A	Ala	aliphatic	115	88.6	-	0.62	
	C	Cys	aliphatic	135	108.5	9.1-9.5	0.68	+
10	D	Asp	aliphatic	150	111.1	4.5 (-)	0.03	+
	E	Glu	aliphatic	190	138.4	4.6 (-)	0.04	+
	F	Phe	aromatic	210	189.9	-	1.00	
15	G	Gly	aliphatic	75	60.1	-	0.50	
	H	His	aliphatic	195	153.2	6.2	0.17	+
	I	Ile	aliphatic	175	166.7	-	0.94	
	K	Lys	aliphatic	200	168.6	10.4 (+)	0.28	+
20	L	Leu	aliphatic	170	166.7	-	0.94	
	M	Met	aliphatic	185	162.9	-	0.74	
	N	Asn	aliphatic	160	114.1	-	0.24	+
25	P	Pro	aliphatic	145	112.7	-	0.71	
	Q	Gln	aliphatic	180	143.8	-	0.25	+
	R	Arg	aliphatic	225	173.4	~ 12 (+)	0.00	+
	-S	Ser	aliphatic	115	89.0		0.36	+
30	T	Thr	aliphatic	140	116.1	-	0.45	+
	V	Val	aliphatic	155	140.0	-	0.83	
	W	Trp	aromatic	255	227.8	-	0.88	+
35	Y	Tyr	aromatic	230	193.6	9.7	0.88	+
	^a Chothia, C., J. Mol. Biol., 105 (1975) 1-14; ^b Zamyatin, A.A., Prog. Biophys. Mol. Biol., 24 (1972) 107-123 ; ^c Tanford, C., Adv. Prot Chem., 17 (1962) 69-165 ; ^d charge at physiological pH; ^e Black, S.D, Mould, D.R, Anal. Biochem., 193 (1991) 72-82.							

[0068] If, for example, the first substrate were ALY and the target substrate were NRF, intermediate substrates with regard to the amino acid composition would be, for example, AL, F, NRY, or ARF (Modifications with regard to the first substrate are indicated). An intermediate substrate with regard to amino acid properties would be, for example, AQF, with the glutamine residue resembling the original leucine in the sense that it is uncharged, but resembling more the arginine residue of the target substrate with respect to hydrophobicity and its capacity to form hydrogen bonds. A further example for this approach would be SLY where S resembles A with respect to volume and surface but is more similar to the target N in terms of hydrophobicity and hydrogen bonding.

[0069] Furthermore, the number of consecutive peptide substrates to be used depends on the relatedness of the first peptide sequence and the target peptide sequence as well as the quantitative specificity of the one or more first proteases. The more unrelated the first peptide sequence and the target peptide sequence are, and the higher the specificity of the one or more first proteases is, the more consecutive intermediate peptide substrates are required.

[0070] In a second variant of this aspect of the invention, different proteases that have specificity for different intermediates are selected in parallel in a first step of the method. In a second step, proteases which have the target specificity are then selected from a population containing randomly recombined chimeras of the proteases selected in the first step. Preferably, the recombination of different proteases selected in parallel is achieved by employing an in-vitro homologous recombination technique, such as the Recombination Chain Reaction described in patent application WO 0134835. Both forms of intermediates can be used for this variant. However, amino acid composition intermediates are preferably employed. Figure 11 shows schematically the basic principle of this variant of the fourth aspect of the invention.

[0071] The different intermediates employed in the first step of this variant are preferably chosen in a way, that the

sum of all modifications introduced into these intermediates equals or resembles the characteristics of the target substrate. As an example, when the first substrate were ALY and the target substrate were NRF, suitable amino acid composition intermediates for this embodiment would be NLY, ARY, and ALE. Proteases having specificity for these three substrates would then be randomly recombined and screened for specificity towards the target substrate of this example NRF.

Other examples, including intermediates who more than one modification are to be constructed analogously.

[0072] In further aspects of the invention, two, three or all four of the different aspects mentioned above are combined with each other. In a preferred combination, screening for proteases with decreased Michaelis-Menten constants is combined with the use of intermediate substrates. In another preferred combination, screening for proteases with decreased Michaelis-Menten constants is combined with the screening of two or more substrates consecutive or in parallel. In a further preferred combination, screening for proteases with decreased Michaelis-Menten constants is combined with using an excess of competing, unlabelled substrate. In a particularly preferred combination, the four aspects, screening for proteases with decreased Michaelis-Menten constants, screening of two or more substrates in parallel, the use of an excess of competing, unlabelled substrate, and the use of intermediate substrates, are combined with each other.

[0073] In a particularly preferred embodiment of the invention the target protease has a specificity similar to tissue-type plasminogen activator and cleaves the target substrate CPGR↓WGG. Such target protease can, among others, be generated by the above defined method of the invention when the starting protease is BAR1 protease from *S. cerevisiae*. In such method the following second/intermediate substrates are preferably utilized:

- (i) WLGLVPGG
- (ii) WLGQVPGG
- (iii) WLGRVPGG
- (iv) WLGRWGG
- (v) CPGRVVGG.

[0074] The sequence-specific protease obtainable by the methods described hereinbefore preferably has a specificity similar to tissue-type plasminogen activator and cleaves the target substrate CPGR↓VVGG. The starting protease preferably is BAR1 protease including, but not limited to, the one depicted in SEQ ID NO:8. Additionally, BAR1 proteases modified by truncation up to 200 aa, preferably in the range of 100 to 200 aa, more preferably in the range of 120 to 180 aa, most preferably in the range of 140 to 160 aa at the C- or N-terminal can be used as starting proteases. Even more preferably the sequence-specific protease is derived from said BAR1 derived protease and has at least one mutation selected from the group comprising the modifications L33I, Y45D, T47A, T59I, N82D, E96V, M107I, N123D, E143D, N151V, I152F, K161E, A163T, T165A, R178S, T221I, E231V, D321N, D367G, M369L, V370I, A399S, K404R and S440L. Particularly preferred among said proteases are those having at least one of D367G, V370I, M107I, I152F, E143D and E231V. The particularly preferred mutants of BAR1 protease may further be modified e.g. by truncation of up to 10 aa at the C- or N-terminal ends thereof or by deletion, insertion or substitution of up to 50 aa, preferably up to 20 aa, most preferably up to 10 aa within its sequence.

Detailed Description of the Figures

[0075] Figure 1 depicts schematically the two alternatives A and B of the method of the invention. Starting with a first protease, the aim of the invention is the generation of an evolved protease with a high specificity for a target peptide substrate which is characterized by its amino acid sequence.

For the purpose of this figure, different shapes represent different amino acid residues, and the inverse profile of the shapes represent the protease's recognition sites, respectively. Shapes with a swung tilde at the top represent any amino acid residue at that position. The active site of the enzyme is indicated by an asterisk, and the arrow indicates the cleavage site within the substrate.

The type of the one or more proteases used as first proteases defines whether alternative A or alternative B is to be employed. In alternative A, the first protease is characterized by an already high specificity towards a defined, first substrate. According to the method of the invention, this specificity has to be changed qualitatively into the target specificity. In alternative B, the first protease has a relatively low specificity, i.e. it does not discriminate between a pool of substrates that differs, for example, at positions P2, P1', and/or P2'. By the process of the invention, only the quantitative specificity of those proteases is increased towards the value of the target specificity.

[0076] Figure 2 distinguishes the two alternatives A and B of the method of the invention by showing schematically the qualitative and quantitative changes in specificity during evolution towards the target specificity. The quantitative specificity *s*, as defined in the framework of this invention, refers to the ratio between all accepted and all possible substrates. The qualitative specificity refers to the amino acid composition and sequence of accepted substrates. Specificities of the first proteases (open circles) and the evolved protease (filled circle) are indicated schematically. In

alternative A, the first protease has a quantitative specificity in the range of the target specificity, but a qualitative specificity that differs from the target specificity. In order to generate the target specificity, the specificity is changed qualitatively only. In alternative B, the first protease has the qualitative specificity of the target substrate, but a quantitative specificity that is far below the target specificity. In order to generate the target specificity, the specificity is changed quantitatively only. When both alternatives are combined, the first protease has neither the qualitative nor the quantitative specificity of the target substrate. In order to generate the target specificity, the specificity is changed quantitatively and qualitatively.

[0077] Figure 3 illustrates schematically how proteases with changed catalytic activities are evolved using the two alternatives A and B of the method of the invention. According to the method of the invention, the catalytic activity (abbreviated as A) can be used as a selection parameter. In alternative A, the first protease hydrolyses only substrate 1, whereas other substrates including the target substrate (T) are not or only very slowly hydrolyzed. By use of the method of the invention, proteases are evolved that hydrolyze specifically the target substrate, whereas other substrates including the first substrate are not or only very slowly hydrolyzed. These evolved proteases are selected by an increase of catalytic activity on the target substrate and a decrease of catalytic activity on the first substrate (comparison approach). Alternatively, selection can be based on the affinity towards the target substrate (affinity approach). In alternative B, the first protease hydrolyzes all substrates including the target substrate (T). By use of the method of the invention, proteases are evolved that hydrolyze specifically the target substrate, whereas other substrates including the first substrate are not or only very slowly hydrolyzed.

These proteases are selected by screening with an excess of competing substrates (competitor approach) or by screening for higher substrate affinity (affinity approach). In general, the evolved protease can be identified by the comparison of the catalytic activity towards offered substrates including the first substrates and the target substrate.

[0078] Figure 4 depicts schematically in two different forms the intermediate approach as one particular aspect of the invention. For description of symbols, refer to figure 1. The intermediate approach uses one or more intermediate substrates to guide the evolution of specificity gradually towards the target specificity in steps as small as necessary. Intermediate substrates are substrates that have an intermediate character when compared with the first and the target substrate. Intermediates can be classified into two forms. First, intermediate substrates can be provided by replacing at least one but less than all amino acid residues of the first substrate with amino acid residues from the target substrate (Intermediate with respect to amino acid composition, Approach 1). Secondly, intermediate substrates can be provided by selectively introducing at defined positions of the substrate amino acid residues whose properties range between those of the corresponding amino acid residue in the first and the target substrate (Intermediate with respect to amino acid properties, Approach 2). As a further alternative, both intermediate approaches can be combined. Preferably, as shown in the figure, the second approach is implemented into the first approach whenever the step between two intermediates is too large.

[0079] Figure 5 illustrates schematically how, according to the invention, proteases with changed catalytic activities are evolved using the intermediate approach. The first protease has a high activity on a first substrate (1) and no or very low activity on all other substrates including the target substrate (T). The following essential step is the provision of an intermediate substrate (2) as illustrated in Figure 4. By screening for catalytic activity on this substrate, protease variants with an increased activity on this intermediate substrate are selected. This intermediate step can be repeated with a gradual variation of the intermediate substrate towards the nature of the target substrate, until an evolved protease is isolated which shows catalytic activity to the target substrate only and no or very low activity on the first substrate and other substrates.

[0080] Figure 6 shows schematically the shuttle vector pPDE that can be used for the method of the invention. The vector comprises a *S. cerevisiae* origin (2 μ ori), an *E. coli* origin (pMB1 ori), a *S. cerevisiae* marker (URA3), an *E. coli* marker (AmpR), and the expression cassette which is composed of a galactose-inducible *S. cerevisiae* promotor (GAL), a signal sequence for secretion of the expressed protein (signal), a KpnI and an XhoI recognition site for inserting the gene of interest, and a terminator (Cyc1).

[0081] Figure 7 shows exemplarily the hydrolysis of a peptide substrate catalyzed by the tobacco etch virus protease monitored by cross-correlation confocal fluorescence spectroscopy (cc-FCS). The peptide substrate with the sequence ENLYFQS is specifically recognized and hydrolyzed by the TEV protease. 100 nM double-labeled peptide (Alexa 488, Cy5) were incubated with (filled squares) and without (open circles) addition of 0.01 U/ μ l protease in assay buffer containing 50 mM Tris-HCl pH 8.0, 0.5 mM EDTA, 10 mM DTT, 0.05% glycerol.

[0082] Figure 8 shows exemplarily a distribution of catalytic activities obtained by screening a population of protease variants on the substrate WLGLVPGG (intermediate 1, see Example VI) using confocal fluorescence spectroscopy. Shown is the frequency N with which a certain catalytic activity (performance, arbitrary units) is identified. Low values represent low catalytic activities, whereas high values represent high catalytic activities on the substrate. Genes encoding variants having highest performance values are isolated and evaluated with respect to their specificity. These variants are then used as first proteases for the next cycle. This procedure is repeated until there are protease variants identified that have the target specificity.

[0083] Figure 9 shows exemplarily the decrease in K_M during evolution toward higher affinity using the affinity approach

of the invention. The protease used as first protease (wild type) in this experiment was subtilisin E from *B. subtilis* which had a K_M of 194 μM . This K_M was gradually decreased by use of the method of the invention by a factor of 7.5 down to 26 μM .

5 [0084] Figure 10 shows exemplarily the change in specificity during evolution of proteases towards the specificity of t-PA. The activity of variants 1, 2, and 3 were evaluated using the substrates intermediate 1, intermediate 2, and intermediate 3 of example VI. The decrease in the substrate concentration corresponds to proteolytic activity. The faster this decrease is, the higher is the catalytic activity of the protease variant. While the first protease has very low activity on intermediate 1, and no activity on intermediates 2 or 3, the evolved variants show various activities on the three intermediate substrates.

10 [0085] Figure 11 depicts schematically a preferred variant of the intermediate approach of the invention (fourth aspect, see below), where proteases are in a first step selected according to their specificity for different intermediate substrates in parallel. Protease are then selected according their specificity for the target substrate from a population containing recombined variants of the protease variants selected in the first step.

15 [0086] Figure 12 shows exemplarily kinetic progression curves for the first protease in comparison with an evolved protease obtained in round 5 of the optimisation method according to the invention. In case of the first substrate the activity of the evolved protease is lower compared to the first protease. This is inverted in case of the 1st and 4th intermediate, where the first protease shows very limited and no turnover of the substrate, respectively.

20 [0087] The invention is further explained by the following Examples. It is understood that the examples and embodiments described therein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to included within the spirit and purview of this application and are considered within the scope of the appended claims. All publications, patents, and patent applications cited herein are hereby incorporated by reference in their entirety for all purposes.

Examples

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[0088] In the following examples, materials and methods of the present invention are provided including the determination of catalytic properties of enzymes obtained by the method. It should be understood that these examples are for illustrative purpose only and are not to be construed as limiting this invention in any manner.

30 [0089] In the experimental examples described below, standard techniques of recombination DNA technology were used that were described in various publications, e.g. Sambrook et al. (1989), Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, or Ausubel et al. (1987), Current Protocols in Molecular Biology 1987-1988, Wiley Interscience, Methods in Yeast Genetics (1994) A Cold Spring Harbour Laboratory Manual, which are incorporated herein in their entirety by reference. Unless otherwise indicated, restriction enzymes, polymerases and other enzymes as well as DNA purification kits were used according to the manufacturers specifications.

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Example I: Molecular cloning of genes encoding protease variants

40 [0090] Genes encoding protease variants were cloned into a vector suitable for extracellular expression of proteins by the yeast *Saccharomyces cerevisiae*. The vector used is a derivate of the plasmid pYES2, which is commercially available from Invitrogen, Inc. A map of the plasmid is shown in Figure 6. The vector contains a 2 μ origin for amplification in *S. cerevisiae*, a pMB1 origin for amplification in *E. coli*, a URA marker for selection in *S. cerevisiae*, a ampicillin resistance marker for selection in *E. coli*, as well as a GAL promoter and a Cyc1 transcription terminator for inducible expression in *S. cerevisiae*. A 90 bp fragment that contains the leader sequence encoding the signal peptide from the BAR1 gene of *S. cerevisiae* was introduced behind the GAL1 promoter. Restriction sites KpnI and XhoI served as insertion sites for heterologous genes to be expressed: Cloning of genes encoding protease variants was done as follows: the coding sequence of the mature protein was amplified by PCR using primers that introduced a KpnI site at the 5' end and a XhoI site at the 3' end. This PCR fragment was cloned into the appropriate sites of the vector and identity was confirmed by sequencing.

50 Example II: Providing populations of protease variants

[0091] A population of protease variants was provided by random modification of genes encoding proteases with known substrate specificities, followed by expression of the protease variants encoded by these modified genes using *S. cerevisiae* as a suitable host organism. First, genes encoding protease variants with known substrate specificities were PCR amplified under error-prone conditions, essentially as described by Cadwell, R.C and Joyce, G.F. (PCR Methods Appl. 2 (1992) 28-33). Error-prone PCR was done using 30 pmol of each primer, 20 nmol dGTP and dATP, 100 nmol dCTP and dTTP, 20 fmol template, and 5 U Taq DNA polymerase in 10 mM Tris HCl pH 7.6, 50 mM KCl, 7 mM MgCl₂, 0.5 mM MnCl₂, 0.01 % gelatin for 20 cycles of 1 min at 94°C, 1 min at 65°C and 1 min at 72°C. The resulting DNA library

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was purified using the Qiaquick PCR Purification Kit following the suppliers' instructions. PCR products were digested with restriction enzymes *XhoI* and *KpnI* and purified as described in Example I. Afterwards, the PCR products were ligated into the vector which was digested with *XhoI* and *KpnI*, gel-purified and dephosphorylated. The ligation products were transformed into *E. coli*, amplified in LB containing ampicillin as marker, and the plasmids were purified using the Qiaquick Plasmid Purification Kit following the suppliers' instructions. Resulting plasmids were transformed into *S. cerevisiae* cells. Populations of protease variants were provided by inducing expression in the transformed *S. cerevisiae* cells by adding 2% galactose to the medium.

Alternatively, genes encoding protease variants with known substrate specificities were statistically recombined at homologous positions by use of the Recombination Chain Reaction, essentially as described in WO 0134835. PCR products of the genes encoding the protease variants were purified using the Qiaquick PCR Purification Kit following the suppliers' instructions, checked for correct size by agarose gel electrophoresis and mixed together in equimolar amounts. 80 µg of this PCR mix in 150 mM TrisHCL pH 7.6, 6.6 mM MgCl₂ were heated for 5 min at 94 °C and subsequently cooled down to 37 °C at 0.05 °C/sec in order to re-anneal strands and thereby produce heteroduplexes in a stochastic manner. Then, 2.5 U Exonuclease III per µg DNA were added and incubated for 20, 40 or 60 min at 37 °C in order to digest different lengths from both 3' ends of the heteroduplexes. The partly digested PCR products were refilled with 0.6 U Pfu polymerase per µg DNA by incubating for 15 min at 72°C in 0.17 mM dNTPs and Pfu polymerase buffer according to the suppliers' instructions. After performing a single PCR cycle, the resulting DNA was purified using the Qiaquick PCR Purification Kit following the suppliers' instructions, digested with *KpnI* and *XhoI* and ligated into the linearized vector. The ligation products were transformed into *E. coli*, amplified in LB containing ampicillin as marker, and the plasmids were purified using the Qiaquick Plasmid Purification Kit following the suppliers' instructions. Resulting plasmids were transformed into *S. cerevisiae* cells. Populations of protease variants were provided by inducing expression in the transformed *S. cerevisiae* cells by adding 2 % galactose to the medium.

Example III: Providing peptide substrates that resemble the target substrate

[0092] All peptide substrates were synthesized on a peptide synthesizer using the approach of Merrifield et al. (Nature. 207 (1965) 522-523). Peptide substrates that resemble the target substrate were designed by substituting the amino acid residues at one or more positions of the first peptide substrate with the amino acid residues at the one or more positions of the target substrate. Alternatively, the amino acid residues at one or more positions of the first peptide substrate were substituted with amino acid residues that have an intermediate character with respect to the amino acid residues of the first peptide substrate and the amino acid residues of the target peptide substrate. For the determination of the intermediate character of amino acid residues refer to Table III. Marker fluorophores were attached to the peptide substrates either via the amino group of the N-terminus or via the carboxy group of the C-terminus. Alternatively, a cysteine residue was added either at the N-terminus or at the C-terminus of the peptide, and the marker fluorophore was chemically attached to the thiol group of the cysteine residue.

Alexa 488 (Molecular Probes Inc., Oregon, USA) and Cy-5 (Amersham Biosciences Europe GmbH, Freiburg, Germany) were typically used as fluorophore markers. Protease cleavage of the peptide substrate was monitored by cross-correlation FCS (Proc.Natl.Acad.Sci.USA. 95 (1998) 1416-1420). As an example, the cleavage of a peptide substrate that contains the target substrate for tobacco etch virus protease (TEV protease) and has the Alexa 488 fluorophore attached to the C-terminus of the peptide and the Cy-5 fluorophore attached to the N-terminus of the peptide is shown in Figure 7. The TEV protease has already a relatively high specificity ($s = 4.9$, see Table I). Cleavage was done at a peptide concentration of 100 nM by adding 0.01 U/µl TEV protease in assay buffer containing 50 mM Tris-HCl pH 8.0, 0.5 mM EDTA, 10 mM DTT, and 0.05% glycerol.

Example IV: Screening procedure

[0093] In order to identify enzyme variants having the desired substrate specificity, a screening approach based on a confocal fluorescence spectroscopy set-up as disclosed in WO 9416313 was used. Either the cell suspension of a *S. cerevisiae* culture directly, or an aliquot of the cell-free supernatant was used as the sample containing the secreted protease variant. After adding the substrate to the sample and incubation for a certain period of time, the samples were subjected to measurement by confocal fluorescence spectroscopy. If necessary, this procedure was repeated several times in order to measure kinetics of the proteolytic cleavage. Consequently, the samples were ranked according to proteolytic activity, and samples exceeding a certain activity threshold were identified in order to isolate the gene encoding the corresponding protease variant. The distribution of proteolytic activities of protease variants obtained by this procedure is shown in Figure 8.

Example V: Generating sequence-specific proteases with increased affinity towards the target peptide substrate by screening at low substrate concentrations

5 [0094] Protease variants that have an increased affinity towards the target peptide substrate were generated by the method of the invention based on screening at low substrate concentrations. By means of error-prone PCR (according to Cadwell, R.C and Joyce, G.F., PCR Methods Appl. 2 (1992) 28-33), a population of protease variants was generated that is related to the alkaline protease subtilisin E from *Bacillus subtilis*, which has a relatively low specificity ($s = 0.82$). This correlates to the relatively high K_M which is in the range of 150 - 200 μM . The population of protease variants was screened at a complexity of 10^6 variants by confocal fluorescence spectroscopy employing substrate concentrations in 10 the range of 10 nM. Variants isolated in this first screen were used as first proteases in a second cycle to provide another population of protease variants. Analogously, variants isolated in subsequent cycles were used as first proteases in the following cycle. The population of variants provided in the second cycle and all subsequent cycles was generated by a combination of error-prone PCR (see above) and in-vitro homologous recombination (according to WO 0134835). Variants isolated from the first four cycles of this procedure were analyzed kinetically. The increase in affinity towards the 15 substrate over the four rounds corresponds to the decrease in K_M of the best performers of each cycle which is shown in Figure 9.

Example VI: Generating sequence-specific proteases with a target substrate specificity resembling the specificity of tissue-type plasminogen activator

20 [0095] Proteases were generated by the method of the invention that had a specificity that was altered towards the specificity of tissue-type plasminogen activator (t-PA). The BAR1 protease from *Saccharomyces cerevisiae* (SEQ ID NO:8) was used as first protease. This protease belongs to the group of aspartic proteinases (Mac Kay et al.; Structure and Function of the Aspartic Proteinases (1991) 161-172). It is specific for peptide substrates containing the amino acid 25 sequence WLQLKPGQ, and catalyses the cleavage at the peptide bond between the second leucine and the lysine residue. Populations of protease variants that were related to the BAR 1 protease or proteases isolated in subsequent screening cycles were generated by means of error-prone PCR (according to Cadwell, R.C and Joyce, G.F., PCR Methods Appl. 2 (1992) 28-33) and in-vitro homologous recombination using the Recombination Chain Reaction (WO 0134835). Protease variants were screened for proteolytic activity at complexities of 10^6 variants by confocal fluorescence 30 spectroscopy. The BAR 1 protease used as first protease already had a relatively high specificity which was in the range of the target specificity. Therefore, a combination of the affinity approach and of the intermediate approach was used. Screening at low concentrations kept the specificity of the protease high, while screening on intermediate substrates enabled the evolution towards the new specificity. Four intermediate substrates were constructed. Intermediate substrate 1 had the amino acid sequence WLGLVPGG, intermediate substrate 2 the amino acid sequence WLGQVPGG, intermediate substrate 3 the amino acid sequence WLGRVPGG, and intermediate four had the sequence WLGRWGG. The 35 target substrate specificity of t-PA is directed to CPGRWGG with cleavage between the arginine residue and the first valine residue. All substrates are shown in Table IV.

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Table IV

		P4	P3	P2	P1	P1'	P2'	P3'	P4'
5	First substrate	W	L	Q	L	K	P	G	Q
	Intermediate 1	W	L	G	L	V	P	G	G
10	Intermediate 2	W	L	G	Q	V	P	G	G
	Intermediate 3	W	L	G	R	V	P	G	G
15	Intermediate 4	W	L	G	R	V	V	G	G
	Target substrate	C	P	G	R	V	V	G	G
20									

[0096] Intermediate 1 was an amino acid composition intermediate due to the fact that it contained at positions P4, P3, P1 and P2' the same amino acid residues as the first substrate, and at positions P2, P1', and P4' the same residues as the target substrate. Intermediate 2 was an amino acid property intermediate with regard to intermediate 1 and the target substrate. It resembled intermediate 1 but contained at position P1 a glutamine residue which has an intermediate character compared to the leucine residue present at that position in the first substrate and the arginine residue present at that position in the target substrate. Intermediate 3 as another amino acid composition intermediate was based on amino acid residues stemming from both, the first substrate and the target substrate, as intermediate substrate 1 does, but, in contrast to the latter one, shared one additional position with the target substrate. Compared to intermediate 3, intermediate 4 shares one further amino acid with the target sequence at position P2'. The changed specificities of different variants that were generated by this method are shown in Figure 10.

[0097] Increase of substrate specificity can also be measured as time-dependent conversion of the substrates, as exemplarily demonstrated in Figure 12. The substrate conversion is presented as the fraction of non-converted substrate over time. As in Figure 12, the first protease and an evolved variant of round 5 differ in their proteolytic activity on the first substrate, intermediate 1 and intermediate 4, respectively. In case of the first substrate the activity of the evolved protease is lower compared to the first protease. This is inverted in case of the 1st and 4th intermediate, where the first protease shows very limited and no turnover of the substrate, respectively, while the evolved protease shows considerable activity on both substrates.

[0098] In this way proteases are generated according to the method of the invention, that have a substrate specificity similar to the human tissue-type plasminogen activator. The proteases generated have at least one mutation at a position out of the group: 33, 45, 47, 59, 82, 96, 107, 123, 143, 151, 152, 161, 163, 165, 178, 221, 231, 321, 367, 369, 370, 399, 404, 440 (based on the numbering of the amino acid sequence of the protease BAR1 listed as SEQ ID NO:8). Preferably, a protease variant evolved from the BAR1 wt protease towards specificity of the human tissue-type plasminogen activator has at least one mutation out of the group: D367G, M369L, V370I, M107I, I152F, E143D, E231V, L33I, Y45D, T47A, T59I, N82D, E96V, N123D, N151V, K161E, A163T, T165A, R178S, T221I, D321N, A399S, K404R and S440L.

[0099] Figure 12 presents the catalytic behaviour of a protease evolved according to the method of the invention in comparison to the starting (first) protease. Starting with BAR1 protease (SEQ ID NO:8) variants are obtained with different mutations. Fig. 12 shows plots reflecting the increase of substrate specificity of a variant of round 5. Investigations done on the amino acid sequence of the exemplified variant of round 5 revealed a particular combination of amino acid substitutions (with numbering equivalent to the numbering of Bar1 protease) as Y45D, T47A, N82D, M107I, E143D, I152F, T165A, E231V, D367G, V370I.

SEQUENCE LISTING

[0100]

<110> DIREVO Biotech AG

<120> Process for Generating Sequence-Specific Proteases by Directed Evolution and Use Thereof

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10 <170> Patent In Ver. 2.1

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15 <213> Artificial Sequence

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<213> Artificial Sequence

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1 5

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<223> Description of Artificial Sequence: Protease substrate

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Trp Leu Gly Arg Val Pro Gly Gly
1 5

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<210> 5

<211> 8

<212> PRT

<213> Artificial Sequence

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<223> Description of Artificial Sequence: Protease substrate

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1 5

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<211> 8

c212> PRT

c213> Artificial Sequence

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<223> Description of Artificial Sequence: Protease substrate

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Cys Pro Gly Arg Val Val Gly Gly
1 5

40

<210> 7

$\langle 211 \rangle$ 7

<212> PRT

<213> Artificial Sequence

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<220>

<223> Description of Artificial Sequence : TEV protease substrate

<400> 7

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31u Asn Leu Tyr Phe Gln Ser
1 5

55

<210> 8

<211> 587

<212> PRT

<213> *Saccharomyces cerevisiae*

<400> 8

	Met	Ser	Ala	Ile	Asn	His	Leu	Cys	Leu	Lys	Leu	Ile	Leu	Ala	Ser	Phe
5	1				5				10					15		
	Ala	Ile	Ile	Asn	Thr	Ile	Thr	Ala	Leu	Thr	Asn	Asp	Gly	Thr	Gly	His
				20					25				30			

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	Leu	Glu	Phe	Leu	Leu	Gln	His	Glu	Glu	Glu	Met	Tyr	Tyr	Ala	Thr	Thr	
			35					40					45				
5	Leu	Asp	Ile	Gly	Thr	Pro	Ser	Gln	Ser	Leu	Thr	Val	Leu	Phe	Asp	Thr	
		50					55					60					
	Gly	Ser	Ala	Asp	Phe	Trp	Val	Met	Asp	Ser	Ser	Asn	Pro	Phe	Cys	Leu	
	65					70					75					80	
10	Pro	Asn	Ser	Asn	Thr	Ser	Ser	Tyr	Ser	Asn	Ala	Thr	Tyr	Asn	Gly	Glu	
					85					90					95		
	Glu	Val	Lys	Pro	Ser	Ile	Asp	Cys	Arg	Ser	Met	Ser	Thr	Tyr	Asn	Glu	
				100					105					110			
15	His	Arg	Ser	Ser	Thr	Tyr	Gln	Tyr	Leu	Glu	Asn	Gly	Arg	Phe	Tyr	Ile	
			115					120					125				
	Thr	Tyr	Ala	Asp	Gly	Thr	Phe	Ala	Asp	Gly	Ser	Trp	Gly	Thr	Glu	Thr	
		130					135					140					
20	Val	Ser	Ile	Asn	Gly	Ile	Asp	Ile	Pro	Asn	Ile	Gln	Phe	Gly	Val	Ala	
	145					150					155					160	
	Lys	Tyr	Ala	Thr	Thr	Pro	Val	Ser	Gly	Val	Leu	Gly	Ile	Gly	Phe	Pro	
					165					170					175		
25	Arg	Arg	Glu	Ser	Val	Lys	Gly	Tyr	Glu	Gly	Ala	Pro	Asn	Glu	Tyr	Tyr	
				180					185					190			
	Pro	Asn	Phe	Pro	Gln	Ile	Leu	Lys	Ser	Glu	Lys	Ile	Ile	Asp	Val	Val	
			195					200					205				
30	Ala	Tyr	Ser	Leu	Phe	Leu	Asn	Ser	Pro	Asp	Ser	Gly	Thr	Gly	Ser	Ile	
		210					215					220					
	Val	Phe	Gly	Ala	Ile	Asp	Glu	Ser	Lys	Phe	Ser	Gly	Asp	Leu	Phe	Thr	
	225					230					235					240	
	Phe	Pro	Met	Val	Asn	Glu	Tyr	Pro	Thr	Ile	Val	Asp	Ala	Pro	Ala	Thr	
					245					250					255		
40	Leu	Ala	Met	Thr	Ile	Gln	Gly	Leu	Gly	Ala	Gln	Asn	Lys	Ser	Ser	Cys	
				260					265					270			
	Glu	His	Glu	Thr	Phe	Thr	Thr	Thr	Lys	Tyr	Pro	Val	Leu	Leu	Asp	Ser	
			275					280					285				
45	Gly	Thr	Ser	Leu	Leu	Asn	Ala	Pro	Lys	Val	Ile	Ala	Asp	Lys	Met	Ala	
						295						300					
	Ser	Phe	Val	Asn	Ala	Ser	Tyr	Ser	Glu	Glu	Glu	Gly	Ile	Tyr	Ile	Leu	
	305					310					315					320	
50	Asp	Cys	Pro	Val	Ser	Val	Gly	Asp	Val	Glu	Tyr	Asn	Phe	Asp	Phe	Gly	
					325					330					335		
	Asp	Leu	Gln	Ile	Ser	Val	Pro	Leu	Ser	Ser	Leu	Ile	Leu	Ser	Pro	Glu	
				340					345					350			
55	Thr	Glu	Gly	Ser	Tyr	Cys	Gly	Phe	Ala	Val	Gln	Pro	Thr	Asn	Asp	Ser	
			355					360					365				

Met Val Leu Gly Asp Val Phe Leu Ser Ser Ala Tyr Val Val Phe Asp
 370 375 380
 5
 Leu Asp Asn Tyr Lys Ile Ser Leu Ala Gln Ala Asn Trp Asn Ala Ser
 385 390 395 400
 10
 Glu Val Ser Lys Lys Leu Val Asn Ile Gln Thr Asp Gly Ser Ile Ser
 405 410 415
 Gly Ala Lys Ile Ala Thr Ala Glu Pro Trp Ser Thr Asn Glu Pro Phe
 420 425 430
 15
 Thr Val Thr Ser Asp Ile Tyr Ser Ser Thr Gly Cys Lys Ser Arg Pro
 435 440 445
 Phe Leu Gln Ser Ser Thr Ala Ser Ser Leu Ile Ala Glu Thr Asn Val
 450 455 460
 20
 Gln Ser Arg Asn Cys Ser Thr Lys Met Pro Gly Thr Arg Ser Thr Thr
 465 470 475 480
 Val Leu Ser Lys Pro Thr Gln Asn Ser Ala Met His Gln Ser Thr Gly
 485 490 495
 25
 Ala Val Thr Gln Thr Ser Asn Glu Thr Lys Leu Glu Leu Ser Ser Thr
 500 505 510
 Met Ala Asn Ser Gly Ser Val Ser Leu Pro Thr Ser Asn Ser Ile Asp
 515 520 525
 30
 Lys Glu Phe Glu His Ser Lys Ser Gln Thr Thr Ser Asp Pro Ser Val
 530 535 540
 35
 Ala Glu His Ser Thr Phe Asn Gln Thr Phe Val His Glu Thr Lys Tyr
 545 550 555 560
 Arg Pro Thr His Lys Thr Val Ile Thr Glu Thr Val Thr Lys Tyr Ser
 565 570 575
 40
 Thr Val Leu Ile Asn Val Cys Lys Pro Thr Tyr
 580 585
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Claims

1. A method for identifying sequence-specific proteases with target substrate specificities which comprises the following steps

 - (a) providing a population of proteases comprised of variants of one first protease or of variants or chimeras of two or more first proteases, said first proteases having a substrate specificity for a particular amino acid sequence of a first peptide substrate;
 - (b) contacting said population of proteases with one or more second substrates, comprising at least one specific amino acid sequence being identical to the target peptide substrate or having an intermediate character with regard to the first substrate and the target substrate but being not present within the first peptide substrate; and
 - (c) selecting one or more protease variants from the population of proteases provided in step (a) having specificity

for said specific amino acid sequence of the second substrates provided in step (b) under conditions that allow identification of proteases that recognize and hydrolyse preferably said specific one amino acid sequence within the second substrates, wherein the screening for protease activity is achieved by adding in excess peptides other than the second peptide, thereby using the added peptides as competitors.

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2. The method of claim 1, wherein the selection conditions in step (c) further comprise

(i) screening for protease activity under low substrate concentrations, thereby increasing affinity for the second substrate, and/or

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(ii) screening for protease activity by using two or more substrates in comparison, thereby increasing the selectivity of the enzyme.

3. The method of claim 1 or 2, wherein steps (a) to (c) are repeated cyclically until one or more protease variants with specificity for the second substrate are identified, and wherein protease variants selected in one cycle are used as first proteases in the following cycle, and wherein at least one cycle and less than 100 cycles are performed.

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4. The method according to any one of claims 1 to 3, wherein only one second substrate is used in the one or more cycles, and wherein the second substrate is identical with the target substrate.

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5. The method according to any one of claims 1 to 3, wherein different second substrates are used, and wherein the second substrates have an intermediate character with regard to the first substrate and the target substrate, and wherein the second substrate that is used in the last cycle is identical with the target substrate.

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6. The method of claim 5, wherein different second substrates are used in consecutive cycles, and wherein each second substrate has intermediate character with regard to the second substrate used before and the target substrate.

7. The method of claim 5 or 6, whereby in at least one cycle steps (b) to (c) are executed with different second substrates in parallel, and wherein the protease variants isolated in such a parallel way are combined and used as first proteases in the next cycle.

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8. The method according to any of claim 5 to 7, wherein the intermediate character of the intermediate substrates is based on

(i) the amino acid composition,

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(ii) the amino acid sequence,

(iii) the physical and/or the chemical properties of the amino acid residues within the specific amino acid sequence, whereby preferably one or more properties from the group consisting of the following amino acid properties is used: the surface, the volume, the isoelectric point, the side chain pKa, the charge, the polarity, the hydrophobicity, or

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(iv) any combination thereof.

9. The method according to any one of claims 1 to 5, wherein the second substrates differ from the first substrates in that 1 to 5 amino acid residues within the specific amino acid sequence are exchanged.

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10. The method according to any one of claims 1 to 9, wherein the second substrates carry functional groups that enable the detection of the hydrolysis of the substrate, said functional groups being

(i) one or more fluorophores or chromophores, whose spectroscopic properties change upon hydrolysis of the peptide, whereby screening is performed through determination of the change in spectroscopic properties; or

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(ii) two fluorophores which are distinguishable by their fluorescence properties and which are attached to opposite ends of the second substrate, whereby the screening is performed through confocal fluorescence spectroscopy at fluorophore concentrations below 1 μ M ; or

(iii) two fluorophores which form a fluorescence resonance energy transfer (FRET) pair and which are attached to opposite ends of the second substrate, whereby screening is performed through determination of the decrease

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in the energy transfer between the two fluorophores ; or

(iv) a first and second autofluorescent protein flanking the second substrate, whereby the screening is performed through confocal fluorescence spectroscopy at substrate concentrations below 1 μ M; or

(v) a fluorophore and a quencher molecule which are attached to opposite ends of the second substrate, whereby

screening is performed through determination of the decrease in quenching of the fluorophore; or
 (vi) a fluorophor or a chromophor and a binding moiety which are attached to opposite ends of the second substrate, whereby screening is performed through determination of binding of the binding moiety to a specific binding partner ; or
 5 (vii) a radioactive label and a binding moiety which are attached to opposite ends of the second substrate, whereby screening is performed through use of a scintillation proximity assay; or
 (viii) any combination thereof.

11. The method according to any one of claims 1 to 10, wherein

- 10 (i) the population of proteases is obtained through random nucleic acid mutagenesis, cassette mutagenesis, site-saturation mutagenesis, site-specific or random insertion and/or deletion mutagenesis, homologous *in vitro* recombination, homologous in-vivo recombination, non-homologous recombination, or a combination thereof; and/or
 15 (ii) the expression of the population of proteases is done by use of host cells, preferably from bacterial, yeast, insect, viral or mammalian origin, or is done by use of cell-free protein expression systems, and/or
 (iii) the coupling of protease genotype and phenotype is achieved by use of sample carriers that enable compartmentation of samples, and the distribution of genotypes into sample carriers is done at a multiplicity per compartment that allows sufficient differentiation of phenotypes.

12. A method according to any of claims 1 to 11, wherein the first protease is selected from the group of proteases consisting of serine proteases, cysteine proteases, aspartic proteases and metalloproteases, and wherein the first protease is preferably selected from the group of proteases consisting of Papain, Bromelain, Trypsin, Pepsin, Chymotrypsin, Subtilisin, SET, Human elastase, Cathepsin, Chymase, *Sacharomycopsis fibuligera* PEP I, Kallikrein, 25 Urokinase, Thermolysin, Collagenase, *Pseudomonas aeruginosa* elastase, TEV protease, HIV-1 protease, BAR1 protease, Factor Xa, Thrombin, Tissue-type plasminogen activator, Kex2 protease, TVMV-protease, RSV protease, MuLV protease, MPMV protease, MMTV protease, BLV protease, EIAV protease, SIVmac protease.

13. The method according to any one of claims 1 to 12, preferably according to any one of claims 5 to 12, wherein the target protease has a specificity similar to tissue-type plasminogen activator and cleaves the target substrate CP-GR↓VVGG.

14. The method of claim 13, wherein the starting protease is BAR1 protease from *S. cerevisiae* and preferably the following second/intermediate substrates are utilized:

- 35 (i) WLGLVPGG
 (ii) WLGQVPGG
 (iii) WLGRVPGG
 (iv) WLGRWGG
 40 (v) CPGRWGG.

Patentansprüche

45 1. Verfahren zur Identifizierung sequenzspezifischer Proteasen mit Zielsubstrat-Spezifitäten, umfassend die folgenden Schritte

- (a) Bereitstellung einer Population von Proteasen, welche Varianten einer ersten Protease oder Varianten oder Chimären von zwei oder mehr ersten Proteasen beinhaltet, wobei diese erste Protease eine Substratspezifität für eine bestimmte Aminosäuresequenz eines ersten Peptidsubstrats aufweist;
 50 (b) Kontaktierung besagter Population von Proteasen mit einem oder mehreren zweiten Substraten, umfassend mindestens eine spezifische Aminosäuresequenz, welche identisch zum Ziel-Peptidsubstrat ist, oder welche einen intermediären Charakter in Bezug auf das erste Substrat und das Zielsubstrat hat, aber nicht im ersten Peptidsubstrat vorhanden ist; und
 55 (c) Selektieren von einer oder mehreren Proteasevarianten aus der Population von Proteasen, bereitgestellt in Schritt (a), mit einer Spezifität für besagte spezifische Aminosäure Sequenz des zweiten Substrats, bereitgestellt in Schritt (b) unter Bedingungen, welche die Identifizierung von Proteasen zulassen, welche bevorzugt besagte spezifische Aminosäuresequenz innerhalb des zweiten Substrats erkennen und hydrolysieren, wobei das Scree-

ning nach Protease-Aktivität durch Zugabe eines Überschusses von anderen Peptiden als dem zweiten Peptid erreicht wird, wobei die zugegebenen Peptide als Kompetitoren dienen.

2. Verfahren nach Anspruch 1, wobei die Selektionsbedingungen in Schritt (c) zudem umfassen
 - (i) Screenen nach Protease-ktivität bei geringen Substratkonzentrationen, wodurch die Affinität für das zweite Substrat erhöht wird, und/oder
 - (ii) Screenen nach Protease Aktivität unter Verwendung von zwei oder mehr Substraten im Vergleich, wodurch die Selektivität des Enzyms erhöht wird.
3. Verfahren nach Anspruch 1 oder 2, wobei die Schritte (a) bis (c) zyklisch wiederholt werden, bis eine oder mehr Proteasevarianten mit Spezifität für das zweite Substrat identifiziert sind, und wobei die in einem Zyklus selektierten Proteasevarianten als erste Proteasen im folgenden Zyklus eingesetzt werden, und wobei mindestens ein Zyklus und weniger als 100 Zyklen ausgeführt werden.
4. Verfahren nach einem der Ansprüche 1 bis 3, wobei nur ein zweites Substrat in dem einen oder mehr Zyklen eingesetzt wird, und wobei das zweite Substrat mit dem Zielsubstrat identisch ist.
5. Verfahren nach einem der Ansprüche 1 bis 3, wobei unterschiedliche zweite Substrate eingesetzt werden, und wobei die zweiten Substrate einen intermediären Charakter in Bezug auf das erste Substrat und das Zielsubstrat haben und wobei das zweite Substrat, das im letzten Zyklus eingesetzt wird, identisch mit dem Zielsubstrat ist.
6. Verfahren nach Anspruch 5, wobei unterschiedliche zweite Substrate in aufeinanderfolgenden Zyklen eingesetzt werden, und wobei jedes zweite Substrat einen intermediären Charakter in Bezug auf das zuvor eingesetzte zweite Substrat und das Zielsubstrat hat.
7. Verfahren nach Anspruch 5 oder 6, wobei in mindestens einem Zyklus Schritte (b) bis (c) mit unterschiedlichen zweiten Substraten parallel ausgeführt werden und wobei die in solch einer parallelen Weise isolierten Protease Varianten kombiniert und als erste Proteasen für den nächsten Zyklus verwendet werden.
8. Verfahren nach einem der Ansprüche 5 bis 7, wobei der intermediäre Charakter des intermediären Substrates basiert auf
 - (i) der Aminosäurezusammensetzung,
 - (ii) der Aminosäuresequenz,
 - (iii) den physikalischen und/oder chemischen Eigenschaften der Aminosäurereste innerhalb der spezifischen Aminosäuresequenz, wobei bevorzugt eine oder mehr Eigenschaften aus der Gruppe bestehend aus den folgenden Aminosäureeigenschaften verwendet wird: die Oberfläche, das Volumen, der isoelektrische Punkt, der Seitenketten pKa, die Ladung, die Polarität, die Hydrophobizität, oder
 - (iv) jegliche Kombination derselben.
9. Verfahren nach einem der Ansprüche 1 bis 5, wobei sich die zweiten Substrate von den ersten Substraten insofern unterscheiden, als dass 1 bis 5 Aminosäurereste innerhalb der spezifischen Aminosäuresequenz ausgetauscht werden.
10. Verfahren nach einem der Ansprüche 1 bis 9, wobei die zweiten Substrate funktionelle Gruppen tragen, welche die Detektion der Hydrolyse des Substrats ermöglichen, wobei besagte funktionelle Gruppen sind
 - (i) ein oder mehr Fluorophore oder Chromophore, deren spektroskopische Eigenschaften sich bei Hydrolyse des Peptids ändert, wodurch das Screening mittels Bestimmung der Änderungen der spektroskopischen Eigenschaften erfolgt; oder
 - (ii) zwei Fluorophore, die durch ihre Fluoreszenzeigenschaften unterscheidbar sind und an die gegenüberliegenden Enden des zweiten Substrats eingebaut sind, wodurch das Screening mittels konfokaler Fluoreszenzspektroskopie bei Fluorophorkonzentrationen unter 1 μ M erfolgt; oder
 - (iii) zwei Fluorophore, die ein Fluoreszenz-Resonanz-Energietransfer(FRET)-Paar bilden, und welche an gegenüberliegenden Enden des zweiten Substrats eingebaut sind, wodurch das Screening durch Bestimmung der Abnahme im Energietransfer zwischen den beiden Fluorophoren erfolgt; oder
 - (iv) ein erstes und zweites autofluoreszierendes Protein, welche das zweite Substrat flankieren, wodurch das

Screening mittels konfokaler Fluoreszenzspektroskopie bei Substratkonzentrationen unter 1 μ M erfolgt; oder
(v) ein Fluorophor und ein Quencher Molekül, welche an gegenüberliegenden Enden des zweiten Substrats eingebaut sind, wodurch das Screening mittels Bestimmung der Abnahme der Quenchung des Fluorophors erfolgt; oder

5 (vi) ein Fluorophor oder ein Chromophor und ein Bindungsrest, welche an gegenüberliegenden Enden des zweiten Substrats eingebaut sind, wodurch das Screening durch Bestimmung der Bindung des Bindungsrests an einen spezifischen Bindungspartner erfolgt; oder

(vii) ein radioaktives Label und ein Bindungsrest, welche an gegenüberliegenden Enden des zweiten Substrats eingebaut sind, wodurch das Screening mittels Anwendung eines Scintillations-Proximity-Assays erfolgt; oder
10 (viii) jegliche Kombination derselben.

11. Verfahren nach einem der Ansprüche 1 bis 10, wobei

15 (i) die Population von Proteasen durch zufallsbestimmte Nukleinsäure Mutagenese, Kassetten-Mutagenese, Orts-Sättigungs-Mutagenese, ortsspezifische oder zufallsbestimmte Insertions- und/oder Deletionsmutagenese, homologe *in-vitro*-Rekombination, homologe *in-vivo*-Rekombination, nichthomologe Rekombination oder einer Kombination derselben erhalten wird; und/oder

(ii) die Expression der Population der Proteasen durch Wirtszellen, bevorzugt durch aus Bakterien, Hefen, Insekten, Viren oder Säugetieren stammenden Wirtszellen, oder durch ein zellfreies Protein-Expressionssystem erfolgt, und/oder

20 (iii) die Kopplung von Protease-Genotyp und -Phänotyp durch den Einsatz von Probenträgern erfolgt, die eine Kompartimentierung der Proben erlauben, und wobei die Verteilung von Genotypen in die Probenträger in einer Vielzahl pro Reaktionsraum erfolgt, die eine ausreichende Differenzierung von Phänotypen erlaubt.

25 12. Verfahren nach einem der Ansprüche 1 bis 11, wobei die erste Protease aus der Gruppe von Proteasen bestehend aus Serin-Proteasen, Cystein-Proteasen, Aspartat-Proteasen und Metalloproteasen ausgewählt wird, und wobei die erste Protease bevorzugt aus der Gruppe von Proteasen, bestehend aus Papain, Bromelain, Trypsin, Pepsin, Chymotrypsin, Subtilisin, SET, humaner Elastase, Kathepsin, Chymase, Saccharomycopsis fibuligera PEP I, Kalikrein, Urokinase, Thermolysin, Kollagenase, Pseudomonas aeruginosa Elastase, TEV-Protease, HIV-1-Protease, BAR1-Protease, Faktor Xa, Thrombin, Tissue-type-PlasminogenAktivator, Kex2-Protease, TVMV-Protease, RSV-Protease, MuLV-Protease, MPMV-Protease, MMTV-Protease, BLV-Protease, EIAV-Protease, SIVmac-Protease ausgewählt wird.

30 13. Verfahren nach einem der Ansprüche 1 bis 12, bevorzugt nach einem der Ansprüche 5 bis 12, wobei die Zielprotease eine ähnliche Spezifität wie Tissue-type-Plasminogen-Aktivator besitzt und das Zielsubstrat CPGR¹VVGG spaltet.

14. Verfahren nach Anspruch 13, wobei die Startprotease die BAR1 Protease aus *S. cerevisiae* ist und bevorzugt folgende sekundären/intermediären Substrate verwendet werden:

- 40 (i) WLGLVPGG
(ii) WLGQVPGG
(iii) WLGRVPGG
(iv) WLGRVVGG
(v) CPGRVVGG.

Revendications

50 1. Procédé d'identification de protéases à séquences spécifiques avec des spécificités de substrat cible qui comprend les étapes suivantes :

(a) fournir une population de protéases composée de variants d'une première protéase ou de variants ou chimères de deux premières protéases ou plus, lesdites premières protéases possédant une spécificité de substrat pour une séquence d'acides aminés particulière d'un premier substrat peptidique ;

55 (b) mettre en contact ladite population de protéases avec un ou plusieurs deuxièmes substrats, comprenant au moins une séquence d'acides aminés spécifique étant identique au substrat peptidique cible ou possédant un caractère intermédiaire par rapport au premier substrat et au substrat cible mais n'étant pas présente à l'intérieur du premier substrat peptidique ; et

- (c) sélectionner un ou plusieurs variants de protéases à partir de la population de protéases fournie à l'étape (a) possédant une spécificité pour ladite séquence d'acides aminés spécifique des deuxièmes substrats fournis à l'étape (b) dans des conditions qui permettent l'identification de protéases qui reconnaissent et hydrolysent de préférence ladite séquence d'acides aminés spécifique à l'intérieur des deuxièmes substrats, dans lequel le criblage pour une activité protéase est réalisé en ajoutant un excès de peptides différents du deuxième peptide, pour utiliser ainsi les peptides ajoutés comme concurrents.
2. Procédé selon la revendication 1, dans lequel les conditions de sélection à l'étape (c) comprennent en outre
- (i) le criblage pour une activité protéase à de faibles concentrations de substrat, pour augmenter ainsi l'affinité pour le deuxième substrat, et/ou
- (ii) le criblage pour une activité protéase en utilisant deux substrats ou plus en comparaison, pour augmenter ainsi la sélectivité de l'enzyme.
3. Procédé selon la revendication 1 ou 2, dans lequel les étapes (a) à (c) sont répétées de façon cyclique jusqu'à ce qu'un ou plusieurs variants de protéases avec une spécificité pour le deuxième substrat soient identifiés, et dans lequel les variants de protéases sélectionnés lors d'un cycle sont utilisés comme premières protéases lors du cycle suivant, et dans lequel au moins un cycle et moins de 100 cycles sont effectués.
4. Procédé selon l'une quelconque des revendications 1 à 3, dans lequel un seul deuxième substrat est utilisé dans les un ou plusieurs cycles, et dans lequel le deuxième substrat est identique au substrat cible.
5. Procédé selon l'une quelconque des revendications 1 à 3, dans lequel différents deuxièmes substrats sont utilisés, et dans lequel les deuxièmes substrats possèdent un caractère intermédiaire par rapport au premier substrat et au substrat cible, et dans lequel le deuxième substrat qui est utilisé lors du dernier cycle est identique au substrat cible.
6. Procédé selon la revendication 5, dans lequel différents deuxièmes substrats sont utilisés lors de cycles consécutifs, et dans lequel chaque deuxième substrat possède un caractère intermédiaire par rapport au deuxième substrat utilisé précédemment et au substrat cible.
7. Procédé selon la revendication 5 ou 6, par lequel lors d'au moins un cycle les étapes (b) à (c) sont exécutées avec différents deuxièmes substrats en parallèle, et dans lequel les variants de protéases isolés dans une telle voie parallèle sont combinés et utilisés comme premières protéases lors du cycle suivant.
8. Procédé selon l'une quelconque des revendications 5 à 7, dans lequel le caractère intermédiaire des substrats intermédiaires est basé sur
- (i) la composition en acides aminés,
- (ii) la séquence d'acides aminés,
- (iii) les propriétés physiques et/ou chimiques des résidus d'acides aminés à l'intérieur de la séquence d'acides aminés spécifique, une ou plusieurs propriétés dans le groupe constitué par les propriétés d'acides aminés suivantes étant de préférence utilisées : la surface, le volume, le point isoélectrique, le pKa des chaînes latérales, la charge, la polarité, l'hydrophobie, ou
- (iv) toute combinaison de celles-ci.
9. Procédé selon l'une quelconque des revendications 1 à 5, dans lequel les deuxièmes substrats diffèrent des premiers substrats en ce que 1 à 5 des résidus d'acides aminés à l'intérieur de la séquence d'acides aminés spécifique sont échangés.
10. Procédé selon l'une quelconque des revendications 1 à 9, dans lequel les deuxièmes substrats portent des groupes fonctionnels qui permettent la détection de l'hydrolyse du substrat, lesdits groupes fonctionnels étant
- (i) un ou plusieurs fluorophores ou chromophores, dont les propriétés spectroscopiques changent lors de l'hydrolyse du peptide, d'où il résulte que le criblage est effectué par détermination du changement des propriétés spectroscopiques ; ou
- (ii) deux fluorophores qui sont différenciables par leurs propriétés de fluorescence et qui sont attachés à des extrémités opposées du deuxième substrat, d'où il résulte que le criblage est effectué par spectroscopie de fluorescence confocale à des concentrations de fluorophores inférieures à 1 μ M ; ou

- (iii) deux fluorophores qui forment une paire de transfert d'énergie par résonance de fluorescence (FRET) et qui sont attachés à des extrémités opposées du deuxième substrat, d'où il résulte que le criblage est effectué par détermination de la réduction du transfert d'énergie entre les deux fluorophores ; ou
- 5 (iv) une première et une deuxième protéines autofluorescentes flanquant le deuxième substrat, d'où il résulte que le criblage est effectué par spectroscopie de fluorescence confocale à des concentrations de substrat inférieures à 1 μ M ; ou
- (v) un fluorophore et une molécule de désactivation qui sont attachés à des extrémités opposées du deuxième substrat, d'où il résulte que le criblage est effectué par détermination de la réduction de la désactivation du fluorophore ; ou
- 10 (vi) un fluorophore ou un chromophore et un groupe de liaison qui sont attachés à des extrémités opposées du deuxième substrat, d'où il résulte que le criblage est effectué par détermination de la liaison du groupe de liaison à un partenaire de liaison spécifique ; ou
- (vii) un marqueur radioactif et un groupe de liaison qui sont attachés à des extrémités opposées du deuxième substrat, d'où il résulte que le criblage est effectué par utilisation d'un dosage par scintillation de proximité ; ou
- 15 (viii) toute combinaison de ceux-ci.

11. Procédé selon l'une quelconque des revendications 1 à 10, dans lequel

- 20 (i) la population de protéases est obtenue par mutagenèse d'acides nucléiques aléatoires, mutagenèse par cassette, mutagenèse par saturation de sites, mutagenèse par insertion et/ou délétion dirigées ou aléatoires, recombinaison homologue *in vivo*, recombinaison non homologue, ou une combinaison de celles-ci ; et/ou
- (ii) l'expression de la population de protéases est assurée par l'utilisation de cellules hôtes, de préférence d'origine bactérienne, de levure, d'insecte, d'origine virale ou de mammifère, ou est assurée par l'utilisation de systèmes d'expression protéiques acellulaires ; et/ou
- 25 (iii) le couplage du génotype et du phénotype des protéases est réalisé par l'utilisation de supports d'échantillons qui permettent le compartimentage des échantillons, et la distribution des génotypes dans les supports d'échantillons est réalisée à une multiplicité par compartiment, ce qui permet une différenciation suffisante des phénotypes.

- 30 12. Procédé selon l'une quelconque des revendications 1 à 11, dans lequel la première protéase est choisie dans le groupe de protéases constitué par les sérine protéases, les cystéine protéases, les protéases aspartiques et les métalloprotéases, et dans lequel la première protéase est de préférence choisie dans le groupe de protéases constitué par la papaïne, la bromélaïne, la trypsine, la pepsine, la chymotrypsine, la subtilisine, SET, l'élastase humaine, la cathepsine, la chymase, PEP I de *Saccharomycopsis fibuligera*, la kallicréine, l'urokinase, la thermolysine, la collagénase, l'élastase de *Pseudomonas aeruginosa*, la protéase TEV, la protéase du VIH-1, la protéase BAR1, le facteur Xa, la thrombine, l'activateur du plasminogène de type tissulaire, la protéase Kex2, la protéase du TVMV, la protéase du RSV, la protéase du MuLV, la protéase du MPMV, la protéase du MMTV, la protéase du BLV, la protéase de l'EIAV, la protéase du SIVmac.
- 35

- 40 13. Procédé selon l'une quelconque des revendications 1 à 12, de préférence selon l'une quelconque des revendications 5 à 12, dans lequel la protéase cible possède une spécificité similaire à l'activateur du plasminogène de type tissulaire et clive le substrat cible CPGR↓VVGG.

- 45 14. Procédé selon la revendication 13, dans lequel la protéase de départ est la protéase BAR1 de *S. cerevisiae* et de préférence les deuxième substrats/substrats intermédiaires suivants sont utilisés :

- (i) WLGLVPGG
- (ii) WLGQVPGG
- (iii) WLGRVPGG
- 50 (iv) WLGRVVGG
- (v) CPGRVVGG

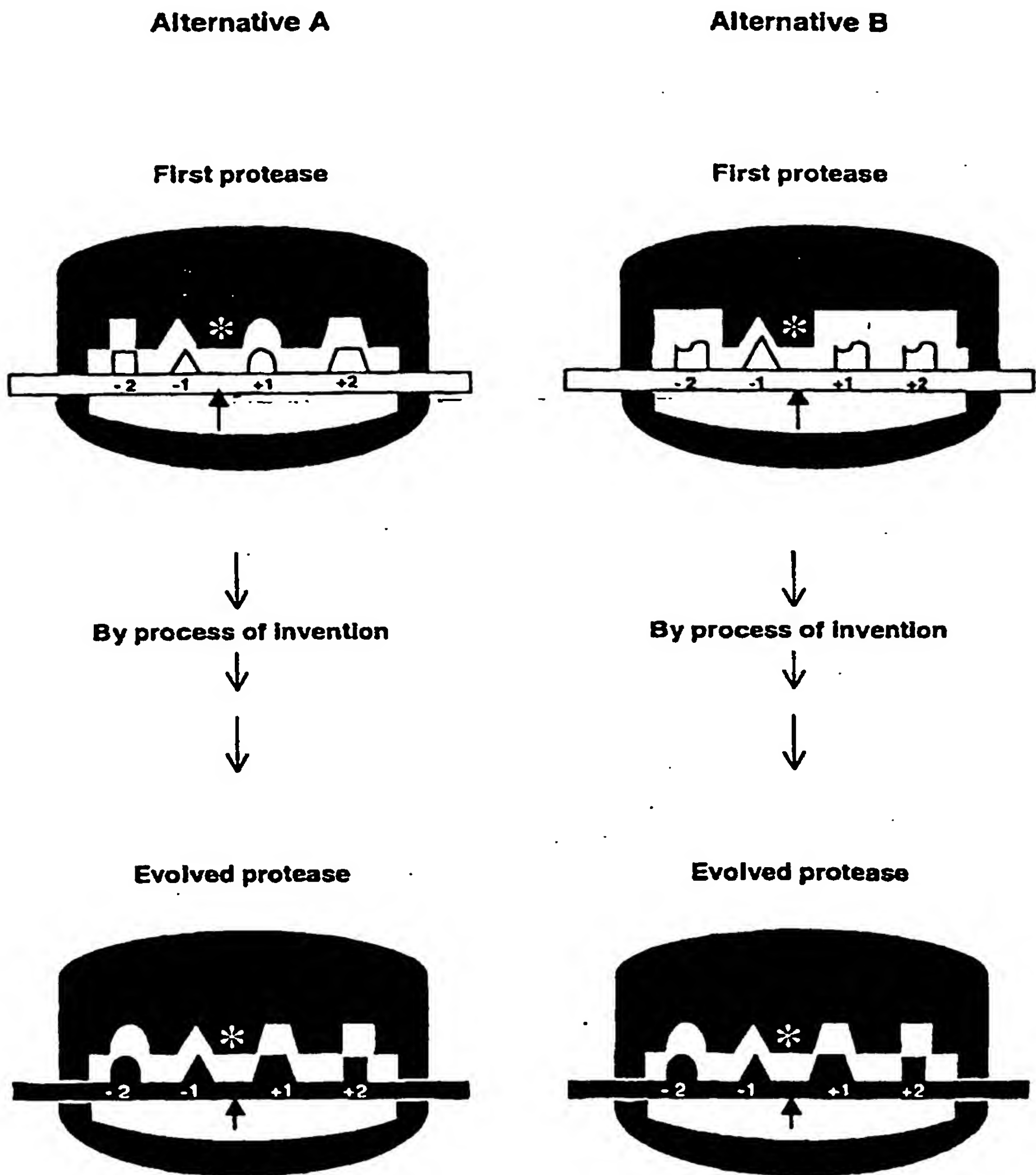


Fig.1

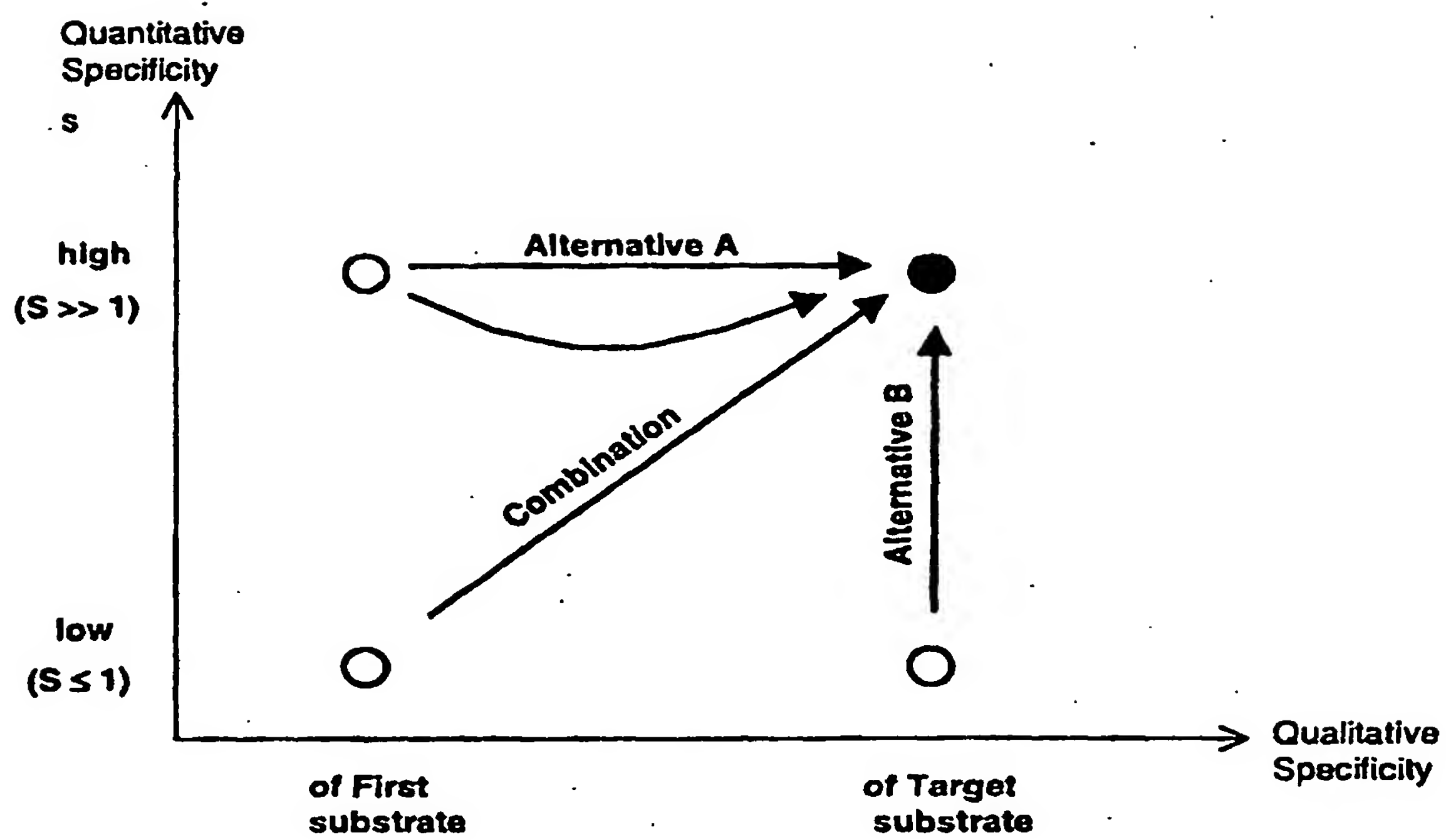


Fig.2

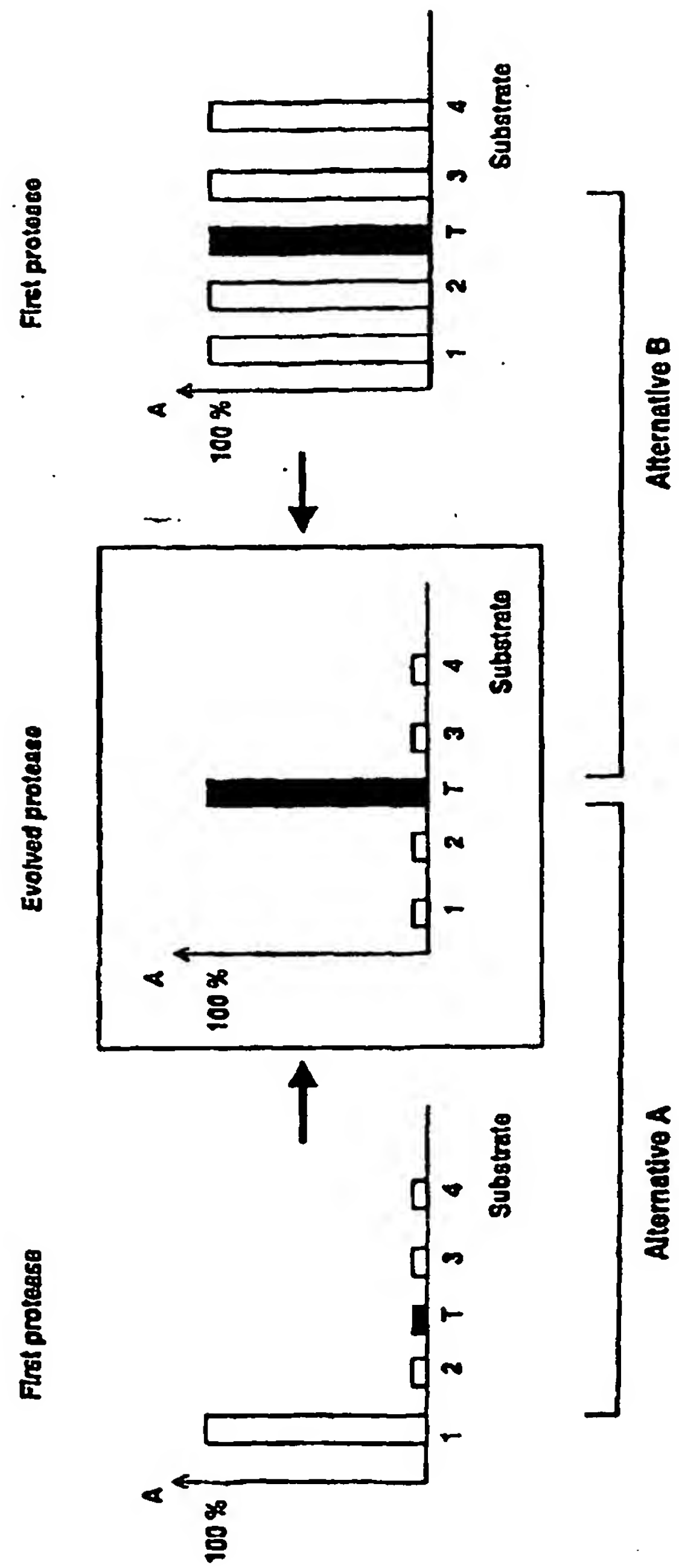


Fig.3

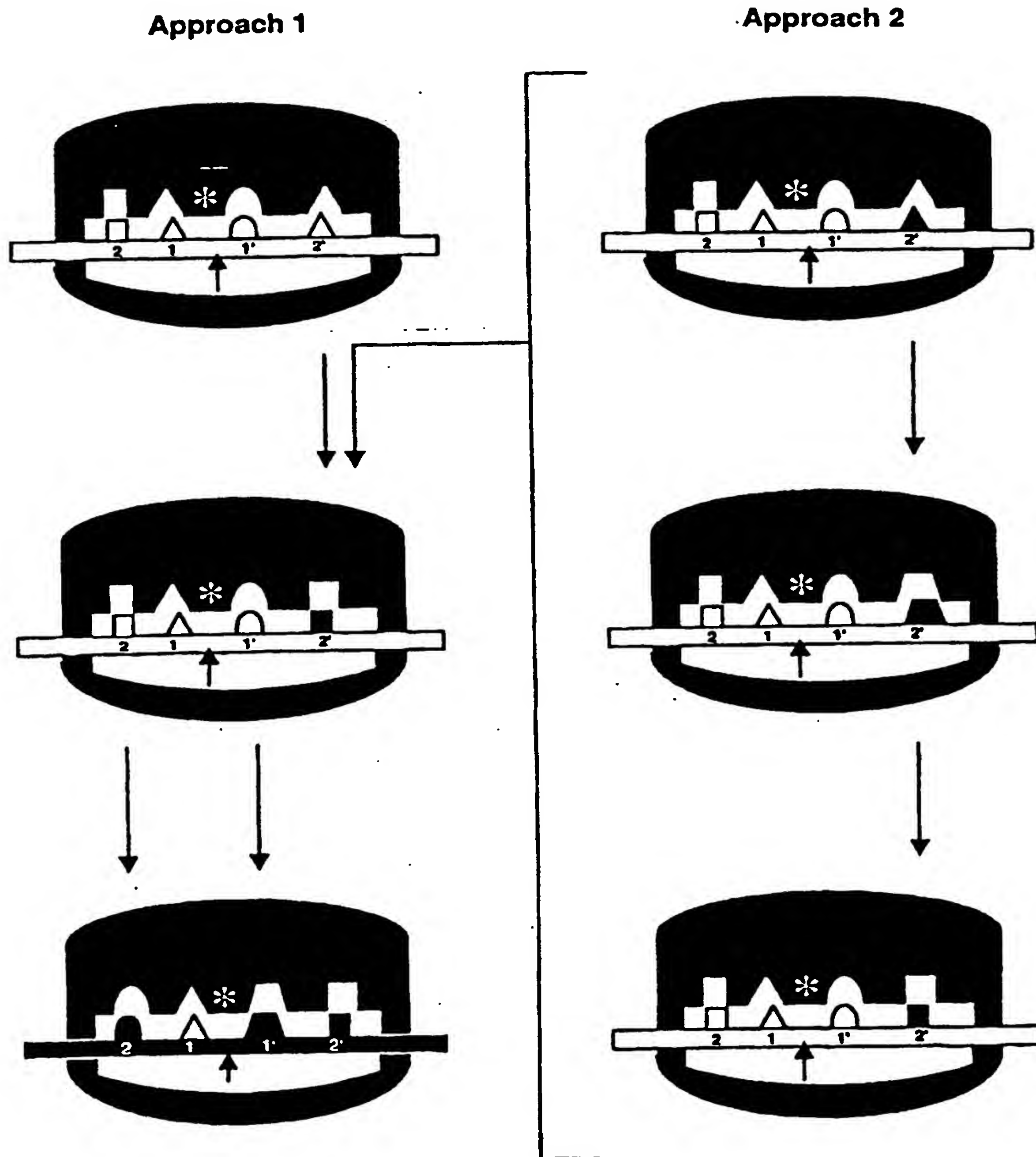


Fig.4

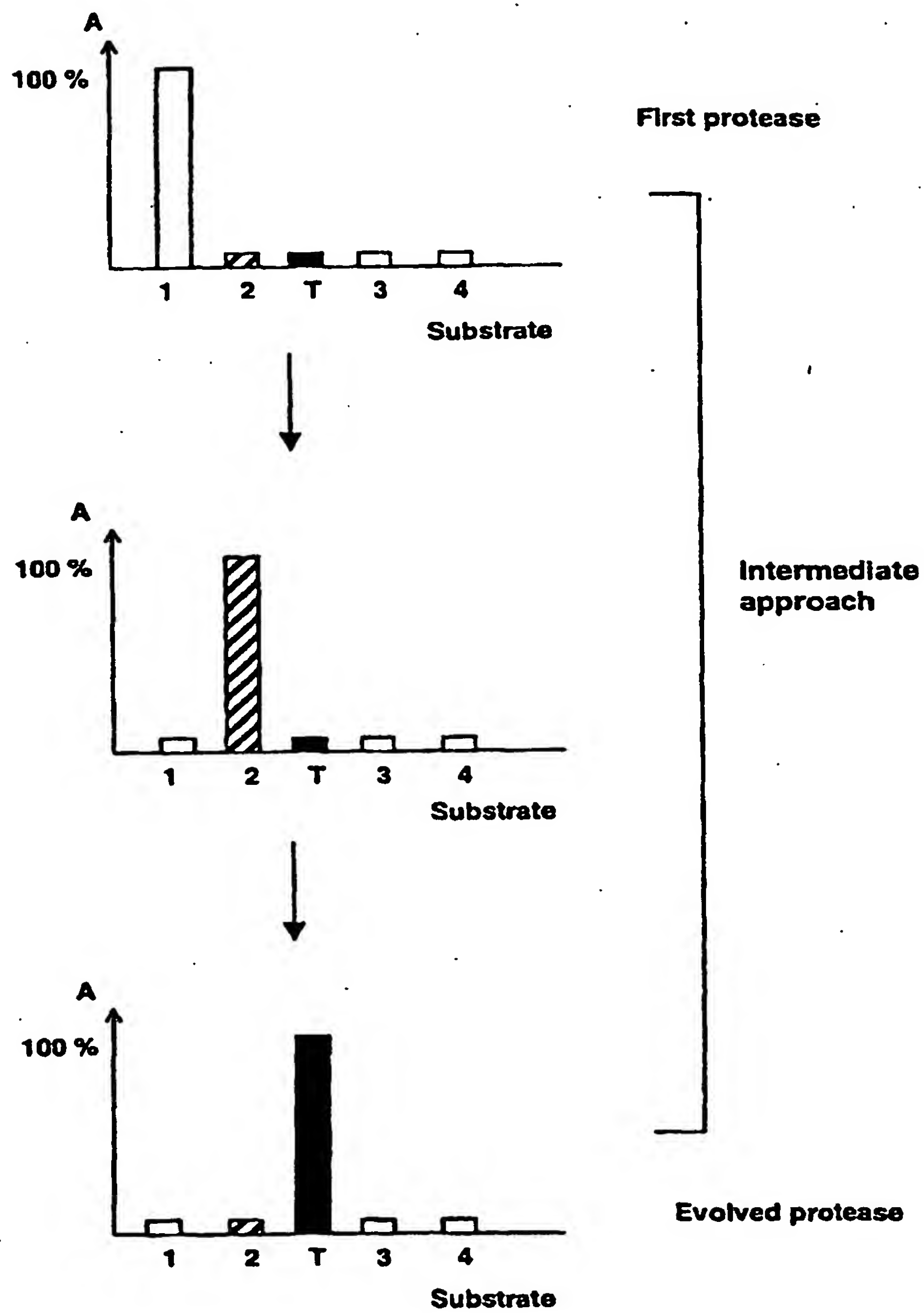


Fig.5

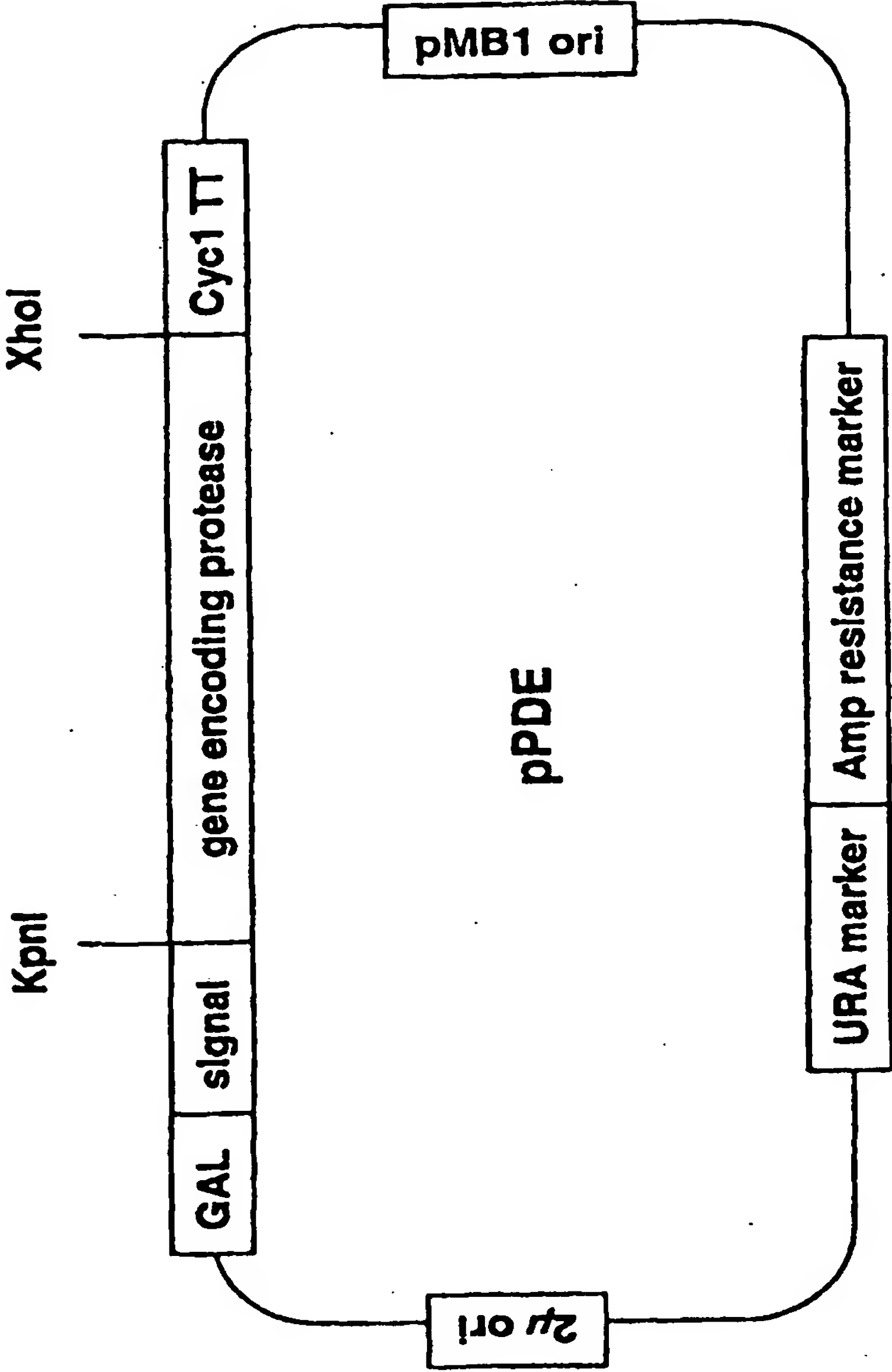
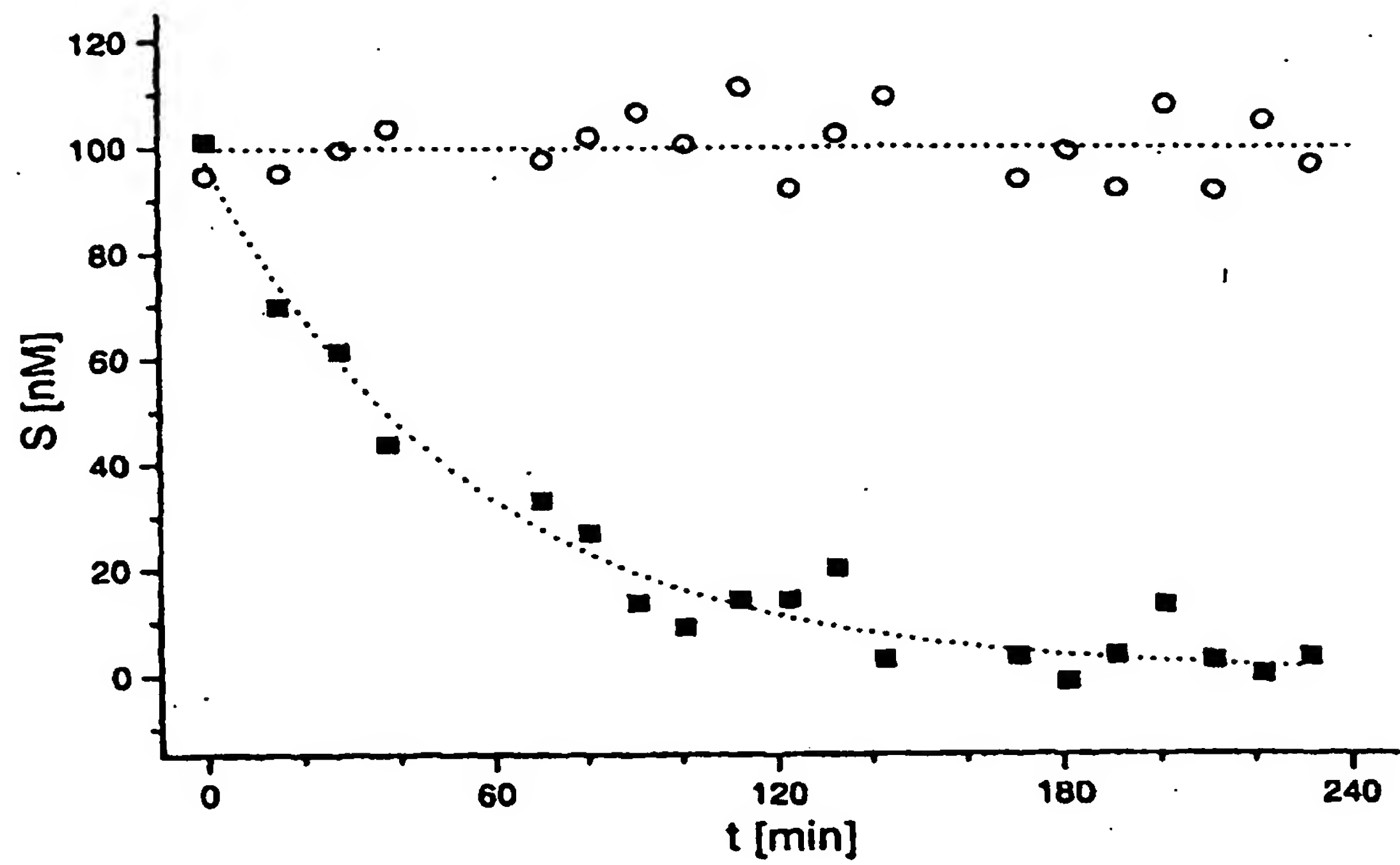
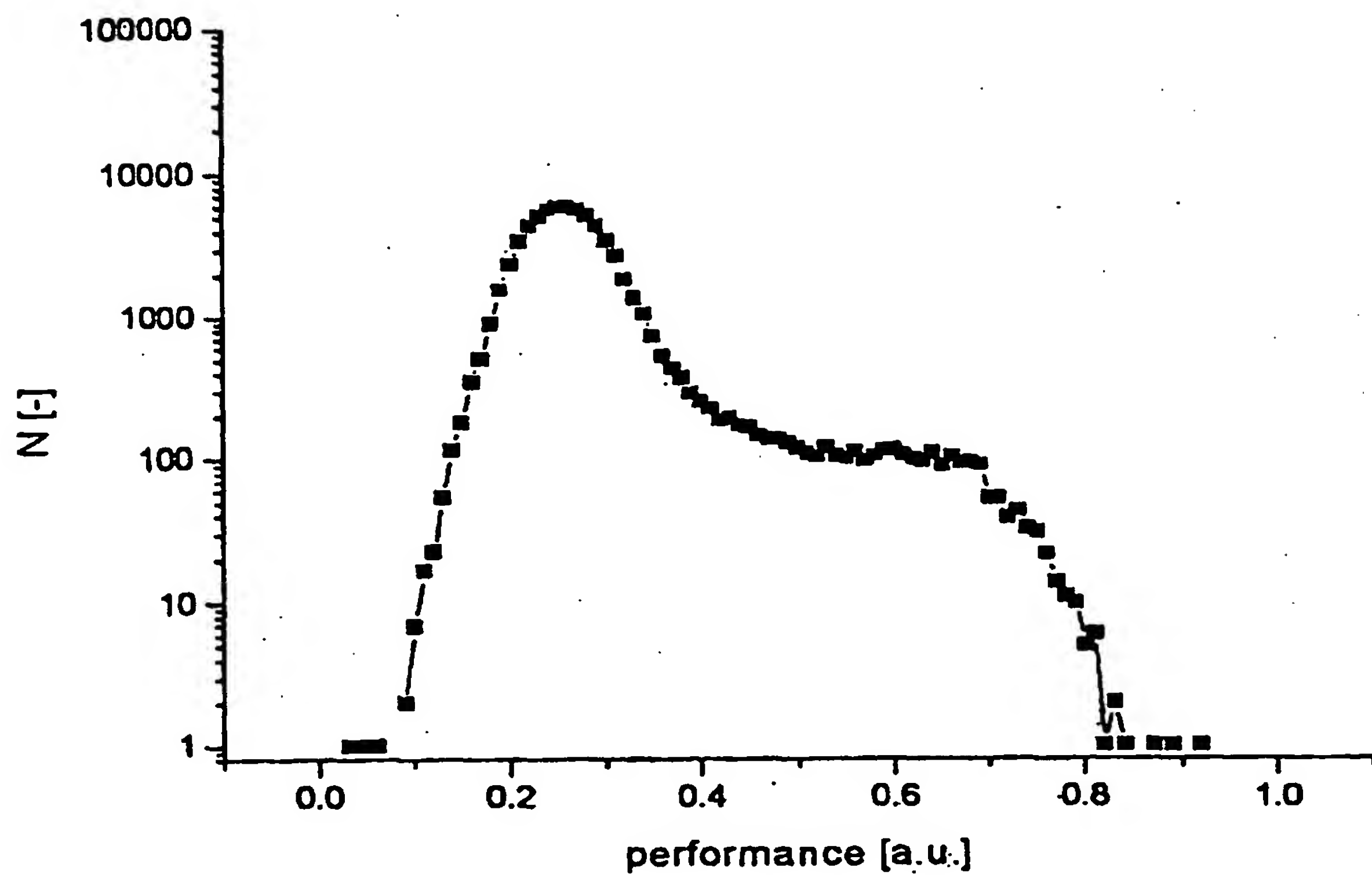
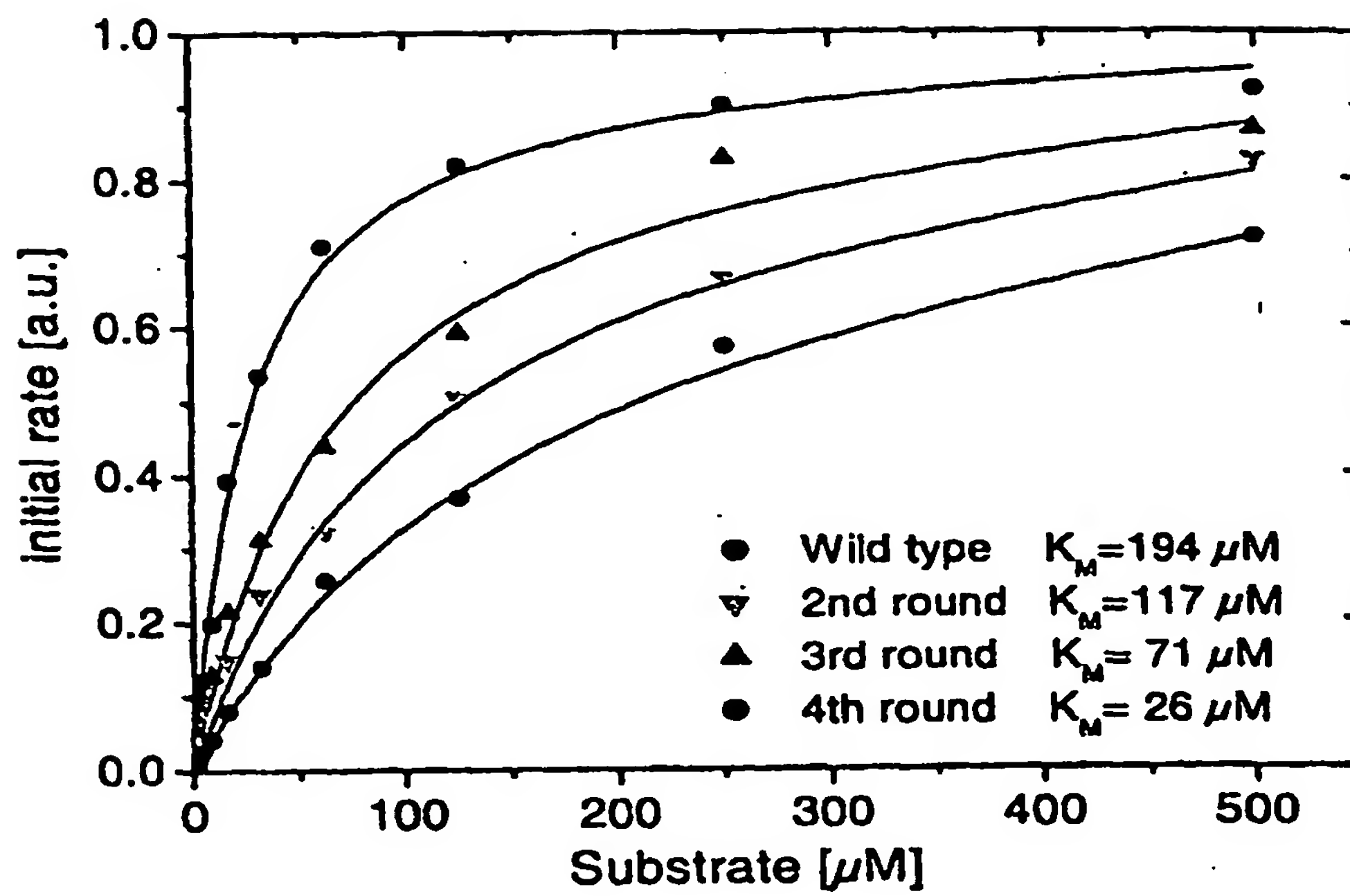


Fig.6

**Fig.7**

**Fig.8**

**Fig.9**

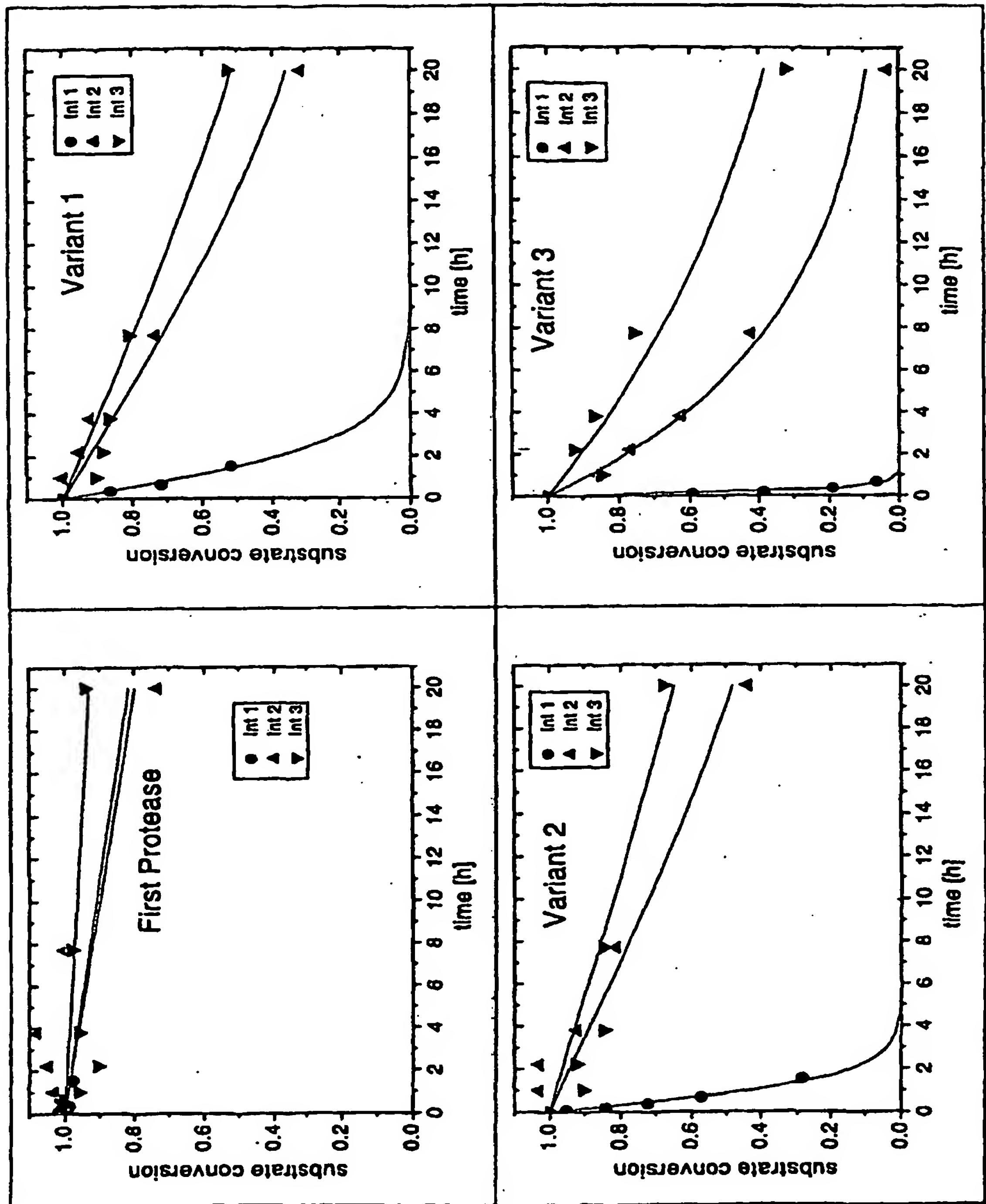
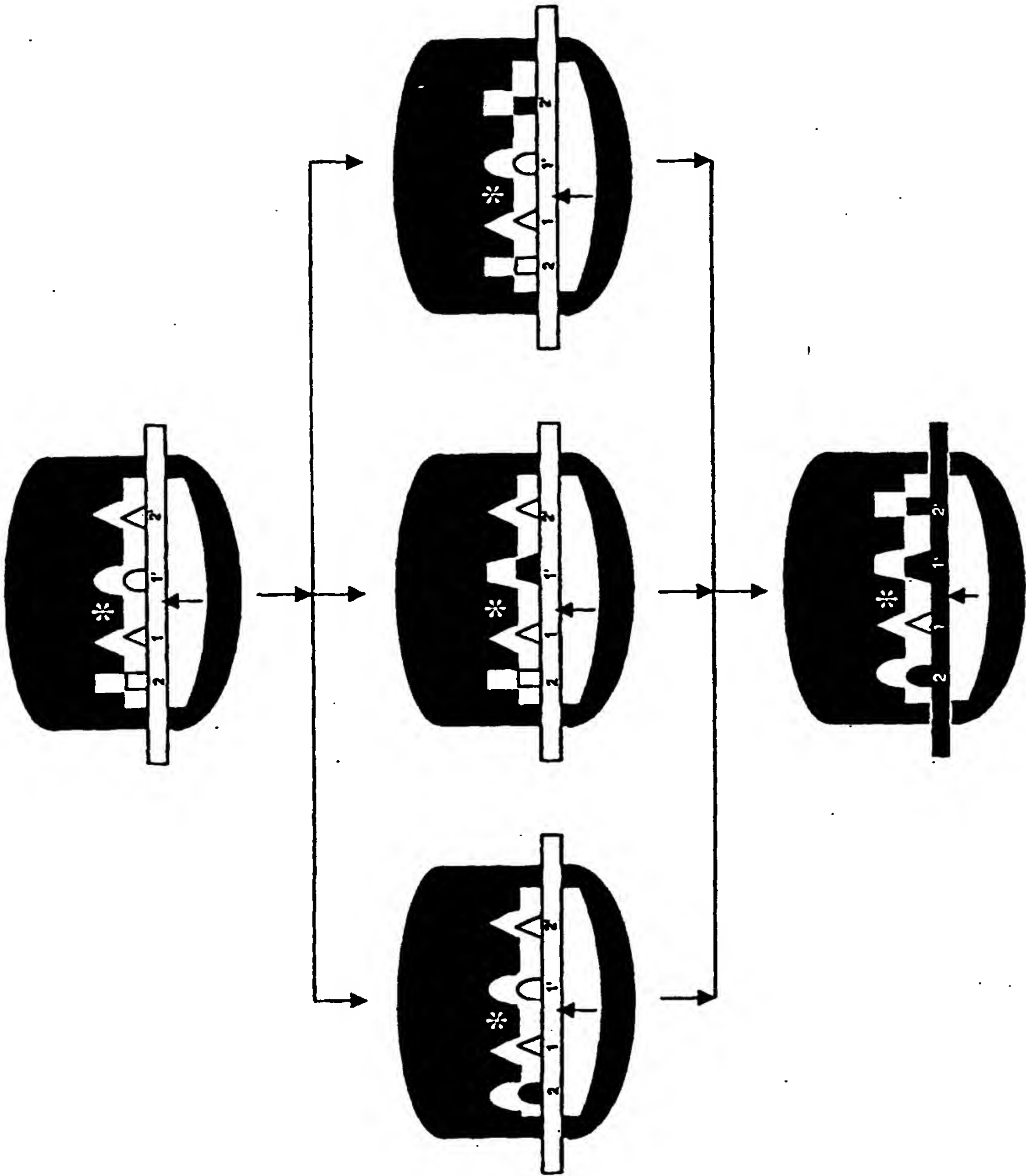
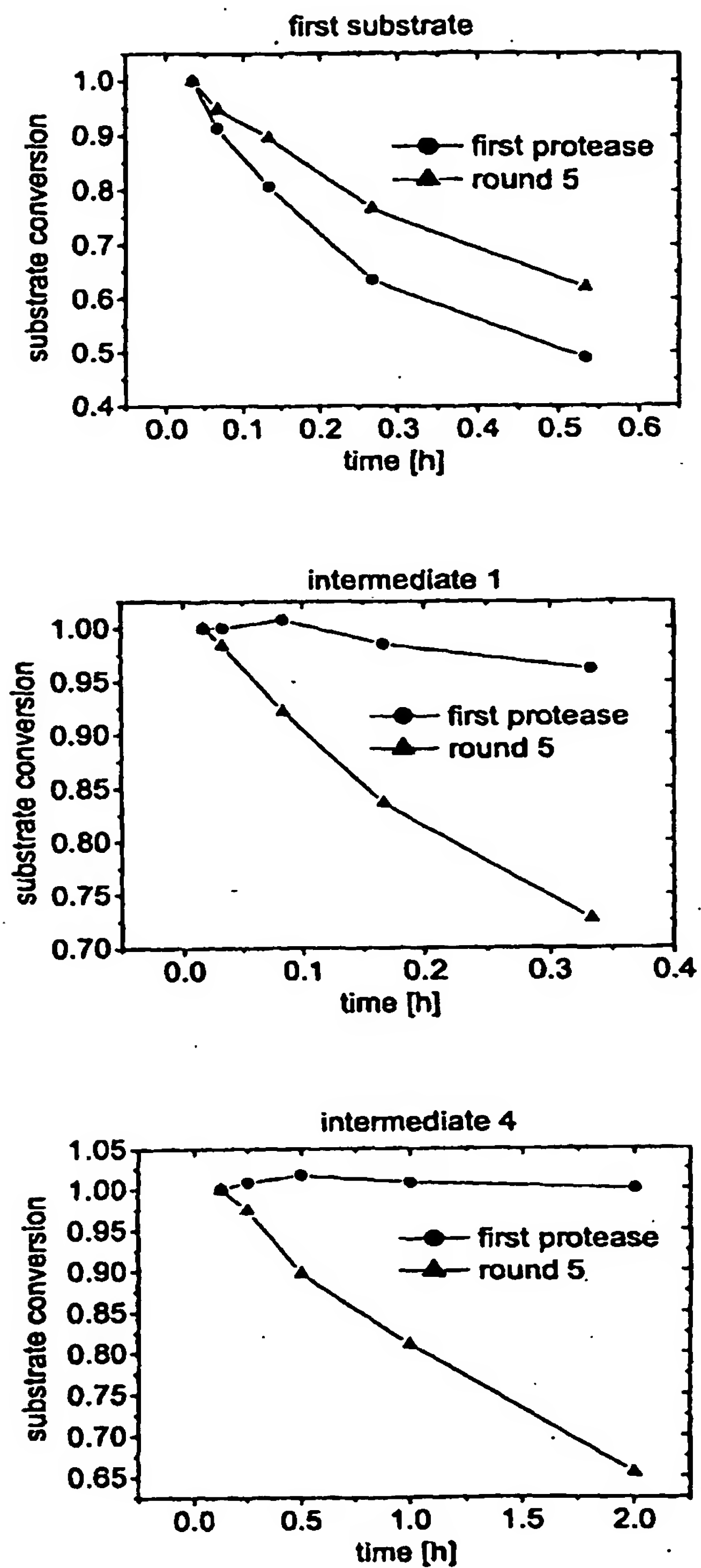


Fig.10



**Fig.12**

REFERENCES CITED IN THE DESCRIPTION

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Patent documents cited in the description

- US 2002022243 A, Harris [0007]
- US 5258289 A [0008]
- WO 9621009 A [0008]
- WO 9849286 A [0009]
- WO 9811237 A, Duff [0010]
- WO 9911801 A, Broad [0011]
- WO 9218645 A [0032] [0037] [0042] [0042]
- WO 0212543 A [0039] [0056] [0061]
- WO 0134835 A [0045] [0070] [0091] [0094] [0095]
- WO 9522625 A [0045]
- WO 9842728 A [0045] [0045]
- WO 9416313 A [0055] [0093]
- WO 9613744 A [0055]

Non-patent literature cited in the description

- SCHLECHTER ; BERGER. *Biochem. Biophys. Res. Commun.*, 1967, vol. 27, 157-162 [0003] [0019] [0066]
- SMITH et al. *Proc. Natl. Acad. Sci. USA*, 1991, vol. 88 [0006]
- KOLTERMANN ; KETTLING. *Biophys. Chem.*, 1997, vol. 66, 159-177 [0033]
- LESLY et al. *Methods in Molecular Biology*, 1995, vol. 37, 265-278 [0041]
- *PCR Methods Appl.*, 1992, vol. 2, 28-33 [0042]
- OLIPHANT, A.R. et al. *Gene*, 1986, vol. 44, 177-183 [0043]
- HORWITZ, M.S. et al. *Genome*, 1989, vol. 31, 112-117 [0043]
- HALLET et al. *Nucleic Acids Res.*, 1997, vol. 25, 1866ff [0044]
- HORTON. *Gene*, 1989, vol. 77, 61ff [0044]
- OSTERMEIER, M. et al. *Nature Biotechnology*, 1999, vol. 17, 1205-1209 [0045]
- CADWELL, R.C ; JOYCE, G.F. *PCR Methods Appl.*, 1992, vol. 2, 28-33 [0046] [0091] [0094] [0095]
- MERRIFIELD et al. *Nature*, 1965, vol. 207, 522-523 [0053] [0092]
- CHOTHIA, C. *J. Mol. Biol.*, 1975, vol. 105, 1-14 [0067]
- ZAMYATIN, A.A. *Prog. Biophys. Mol. Biol.*, 1972, vol. 24, 107-123 [0067]
- TANFORD, C. *Adv. Prot. Chem.*, 1962, vol. 17, 69-165 [0067]
- BLACK, S.D ; MOULD, D.R. *Anal. Biochem.*, 1991, vol. 193, 72-82 [0067]
- SAMBROOK et al. *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor Laboratory, 1989 [0089]
- AUSUBEL et al. *Current Protocols in Molecular Biology* 1987-1988. Wiley Interscience, 1987 [0089]
- *Methods in Yeast Genetics. A Cold Spring Harbour Laboratory Manual*, 1994 [0089]
- *Proc.Natl.Acad.Sci.USA.*, 1998, vol. 95, 1416-1420 [0092]